

Prevalence and Antibiotic Resistance Patterns of Multidrug-Resistant Bacteria Isolated from Clinical Specimens

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Abstract

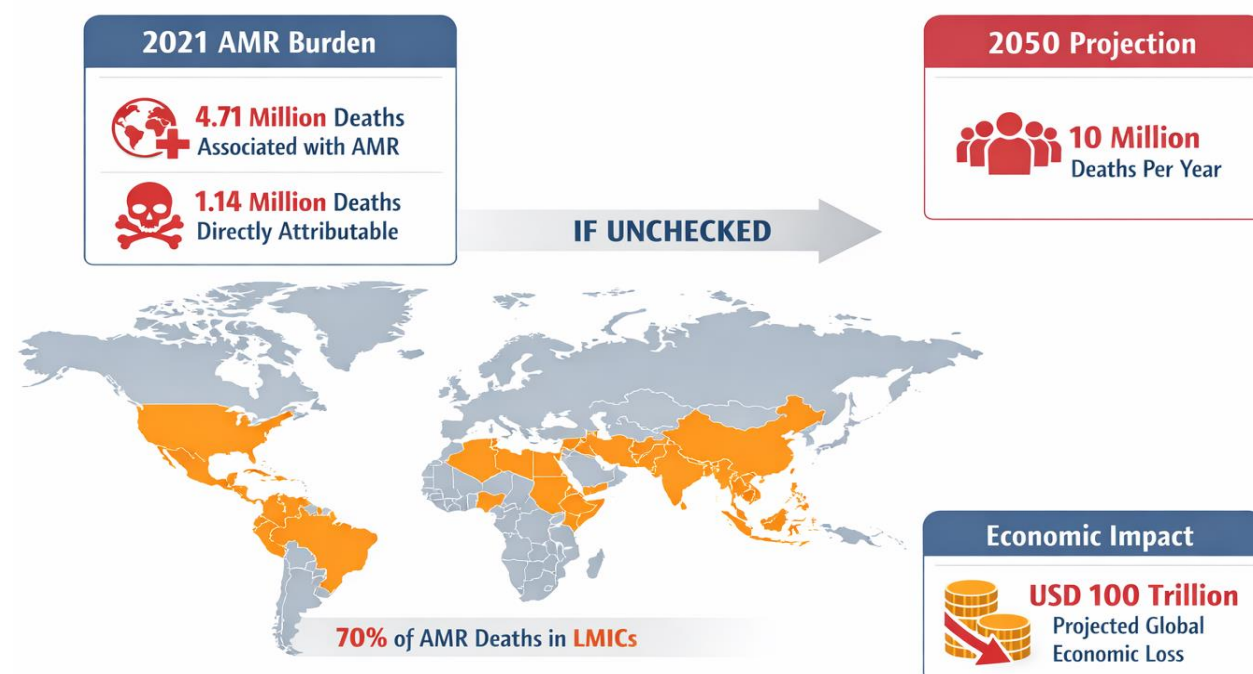
The escalating global crisis of antimicrobial resistance (AMR) poses a severe threat to public health, with multidrug-resistant (MDR) bacteria responsible for an estimated 4.71 million deaths in 2021 and projected to cause 10 million annual deaths by 2050. This review synthesizes current epidemiological data on the prevalence and resistance patterns of MDR bacteria isolated from clinical specimens across diverse geographic regions, with particular emphasis on low- and middle-income countries (LMICs) in Africa and South Asia. High MDR rates were consistently observed across specimen types, with urine (up to 72%), blood (52.3%), and wound/pus samples showing the highest burden. ESKAPE pathogens (*Enterococcus faecium*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, and *Enterobacter* spp.) dominate, exhibiting resistance to multiple antibiotic classes through mechanisms including β -lactamase production (ESBLs, carbapenemases), target modification, efflux pumps, and biofilm formation. Regional hotspots include West Africa (59% overall MDR), Pakistan (high XDR *Salmonella typhi*

and MRSA), and Nepal (52.3% MDR in tertiary care). The analysis underscores the critical role of inappropriate antibiotic use, environmental reservoirs, and healthcare-associated transmission in driving resistance. Strengthening antimicrobial stewardship, rapid diagnostics, enhanced surveillance (e.g., WHO GLASS), and a One Health approach are urgently required to curb the spread of MDR pathogens and preserve the efficacy of existing antimicrobials.

Introduction

The global landscape of clinical medicine is currently defined by a profound and escalating crisis in antimicrobial efficacy, driven by the emergence and dissemination of multidrug-resistant (MDR) bacterial pathogens (Gajic et al., 2025). This phenomenon, which represents a critical inflection point in infectious disease management, is characterized by the evolution of microorganisms that demonstrate non-susceptibility to at least one antimicrobial agent in three or more distinct antibiotic classes (GBD 2021 Antimicrobial Resistance Collaborators, 2024). Recent longitudinal assessments indicate a concerning 43% increase in multidrug-resistant infections on a global scale, with healthcare-associated infections (HAIs) rising by 67% and community-acquired infections increasing by 38% in geographic regions marked by significant antibiotic misuse (Pan et al., 2025). The human and economic tolls of this escalation are staggering; in 2021 alone, bacterial antimicrobial resistance (AMR) was associated with an estimated 4.71 million deaths, with 1.14 million of those deaths directly attributable to resistant pathogens (Karnwal et al., 2025). Projections based on current trajectories suggest that by 2050, the annual global mortality rate attributed to AMR could reach 10 million, with a cumulative economic burden nearing USD 100 trillion if containment strategies are not urgently optimized (Elmubark et al., 2025). The scale of the antimicrobial resistance crisis is illustrated in figure 1, which summarizes current global mortality and future projections associated with drug-resistant infections. The figure highlights the alarming increase in AMR-related deaths and underscores the urgent need for coordinated global interventions.

Figure 1: Global Burden and Projected Impact of Antimicrobial Resistance



Global and Regional Epidemiological Patterns of Resistance

The prevalence of MDR bacteria exhibits marked heterogeneity across different geographical regions, a variation largely dictated by local antibiotic stewardship practices, healthcare infrastructure, and environmental factors (Kashef Hamadani, 2023). While high-income countries face challenges related to the persistence of

resistant clones within specialized hospital units, low- and middle-income countries (LMICs) bear a disproportionate share of the global mortality burden, accounting for approximately 70% of all AMR-related deaths (Sah et al., 2023).

The African Continent: Surveillance Gaps and Escalating Prevalence

In the African region, the prevalence of MDR bacterial infections is an acute threat, with over one million deaths associated with bacterial AMR reported in 2021. In East Africa, research indicates that 50% to 60% of urinary tract infections are now caused by MDR bacteria, while in South Africa, over 80% of clinical isolates of *Acinetobacter baumannii* demonstrate an MDR phenotype (Sartorius et al., 2024). West African meta-analyses involving thirteen countries reveal an overall MDR prevalence of 59%, with significant contributions from Nigeria (34%) and Ghana (22%) (Khasapane et al., 2025). A critical observation in this region is the 7% increase in MDR prevalence recorded over the last decade, with nosocomial infections showing higher resistance rates (65%) compared to community-acquired infections (53%) (Diop et al., 2025).

In Rwanda, the healthcare system struggles with a high prevalence of extended-spectrum beta-lactamase (ESBL) producers. Studies at the University Teaching Hospital of Kigali identified that 75.9% of Gram-negative isolates were resistant to ceftriaxone, and 71.7% exhibited an ESBL-positive phenotype (Osen et al., 2025). This prevalence is directly linked to increased mortality, with deaths in patients infected by ESBL producers being more than double the general hospital mortality rate during the same period (Shamsrizi et al., 2020). Similarly, in Zambia, diagnostic stewardship data from 2020 to 2024 involving over 184,000 specimens show a statistically significant increasing trend in the isolation of priority pathogens like *Escherichia coli*, *Staphylococcus aureus*, and *Klebsiella pneumoniae* (Mudenda et al., 2024).

South Asian Dynamics: The Epicenter of Novel Resistance Mechanisms

South Asia, particularly Pakistan, India, and Nepal, has emerged as a global hotspot for the evolution and rapid dissemination of novel resistance determinants. Pakistan exhibits some of the highest age-standardized mortality rates associated with AMR, with substantial death tolls attributed to *Klebsiella pneumoniae*, *E. coli*, and *S. aureus* (da Silva et al., 2022). A major concern in this region is the emergence of extensively drug-resistant (XDR) *Salmonella typhi*, which has become resistant to almost all first- and second-line treatments (Mirha et al., 2024). Systematic reviews of *S. typhi* in Pakistan reveal alarming resistance to nalidixic acid (92%), ampicillin (80%), and ciprofloxacin (64%), with XDR strains significantly more common in pediatric populations (Ullah et al., 2025).

In Nepal, studies in tertiary care settings show that ESKAPE pathogens (*Enterococcus faecium*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, and *Enterobacter* spp.) are isolated in 32.1% of positive samples, with 52.3% of these being multidrug-resistant (Pandey et al., 2021). In burn units, the situation is even direr, with MDRO prevalence reaching 56.82% and significant predictors of infection including admission to the Intensive Care Unit (ICU) and the use of invasive devices like nasogastric tubes and ventilators (Maharjan et al., 2026).

European and North American Trends

While European data suggests a peak in the prevalence of certain isolates around 2021 followed by some declines, carbapenemase resistance remains high in specific species (Wise et al., 2024). In Palermo, Italy, carbapenemase resistance was observed in 55% of *K. pneumoniae* isolates, although *Pseudomonas aeruginosa* showed a significant decrease in meropenem resistance during the same study period (Pipitò et al., 2025). In the United States, *Enterococcus* species have become the third most frequent

nosocomial pathogen, accounting for 14% of hospital-acquired infections, a significant increase from 11% reported in 2007 (Thomsen et al., 2023).

Table 1: Regional Prevalence of Multidrug-Resistant (MDR) Bacteria and Primary Specimen Sources

Region/Country	MDR Prevalence (%)	Primary Specimen Source	Key Pathogen Trends
West Africa (Meta-analysis)	59%	Urine, Skin Swabs	Rising ESBL in <i>E. coli</i> (Diop et al., 2025)
Sudan (Wad Medani)	39.3%	Blood, Wound Swabs	Higher in males & age groups <5, >60 (Elmubark et al., 2025)
Nepal (Tertiary Care)	52.3%	Urine, Sputum	Predominance of <i>K. pneumoniae</i> (Sah et al., 2023)
India (Udaipur)	61.18% (GNB)	Urine, Pus	High resistance in <i>Klebsiella</i> and <i>E. coli</i> (Pan et al., 2025)
Pakistan (BSI)	29-55% (XDR)	Blood, Bone Marrow	XDR <i>Salmonella</i> and MRSA (Ullah et al., 2025)
Italy (University Hospital)	-	Urine, Respiratory	55% Carbapenem-R <i>K. pneumoniae</i> (Giordano et al., 2023)
China (National Data)	-	Blood, Urine	Fluctuating VREfm (2.2% in 2022) (Zhang et al., 2024)

Clinical Specimen Distribution and Resistance Phenotypes

The prevalence and antibiotic resistance patterns of bacterial isolates vary significantly according to the anatomical site of infection and the type of clinical specimen collected. This variation reflects the unique selective pressures present in different physiological microenvironments (Abebe et al., 2019).

Bloodstream and Systematic Infections

Blood cultures are critical for the identification of pathogens causing sepsis. In studies conducted in Sudan, blood samples demonstrated the highest MDRO detection rate at 52.3%. In Pakistan, *Staphylococcus aureus* (27.9%), *Salmonella* (25%), and *Klebsiella* (13.8%) were the most common isolates from blood cultures, with 88.6% of *S. aureus* being methicillin-resistant (MRSA) (Hoque et al., 2024). The high prevalence of XDR *Salmonella typhi* in blood cultures in South Asia underscores a critical failure of traditional oral and parenteral antibiotic regimens, requiring the use of last-resort carbapenems or azithromycin (Bale, 2021).

Urinary Tract and Catheter-Related Infections

Urinary tract infections (UTIs) represent the most frequent source of clinical isolates globally. In Bangladesh, 47.5% of positive cultures were obtained from urine, with *E. coli* (37.2%) and *Klebsiella* sp. (11.5%) as the dominant pathogens (Bangladesh National Surveillance Team, 2024). In India, urine specimens were the most frequent source of Gram-negative bacilli, showing a 61.18% MDR rate (Pan et al., 2025). Research in West Africa indicates that urine samples exhibit the highest prevalence of MDR bacteria (72%), often associated with ESBL-producing Enterobacteriaceae (Diop et al., 2025). The high rate of resistance to third-generation cephalosporins and

fluoroquinolones in urinary isolates complicates the management of both uncomplicated and catheter-associated UTIs (Sah et al., 2023).

Respiratory and Wound Secretions

Respiratory specimens, including sputum and bronchial lavage, are frequently colonized by *Acinetobacter baumannii* and *Pseudomonas aeruginosa*, particularly in patients undergoing mechanical ventilation (Stamkou et al., 2025). In these samples, *A. baumannii* often demonstrates a difficult-to-treat resistance (DTR) phenotype, with resistance rates for commonly used antibiotics frequently exceeding 80% (Ullah et al., 2025). Wound swabs and pus samples also show high MDR rates, particularly in burn units and surgical wards. In Nepal, *Pseudomonas aeruginosa* (29.0%) and *Klebsiella pneumoniae* (18.1%) were the most common isolates from burn wounds (Ugochi Grace et al., 2025).

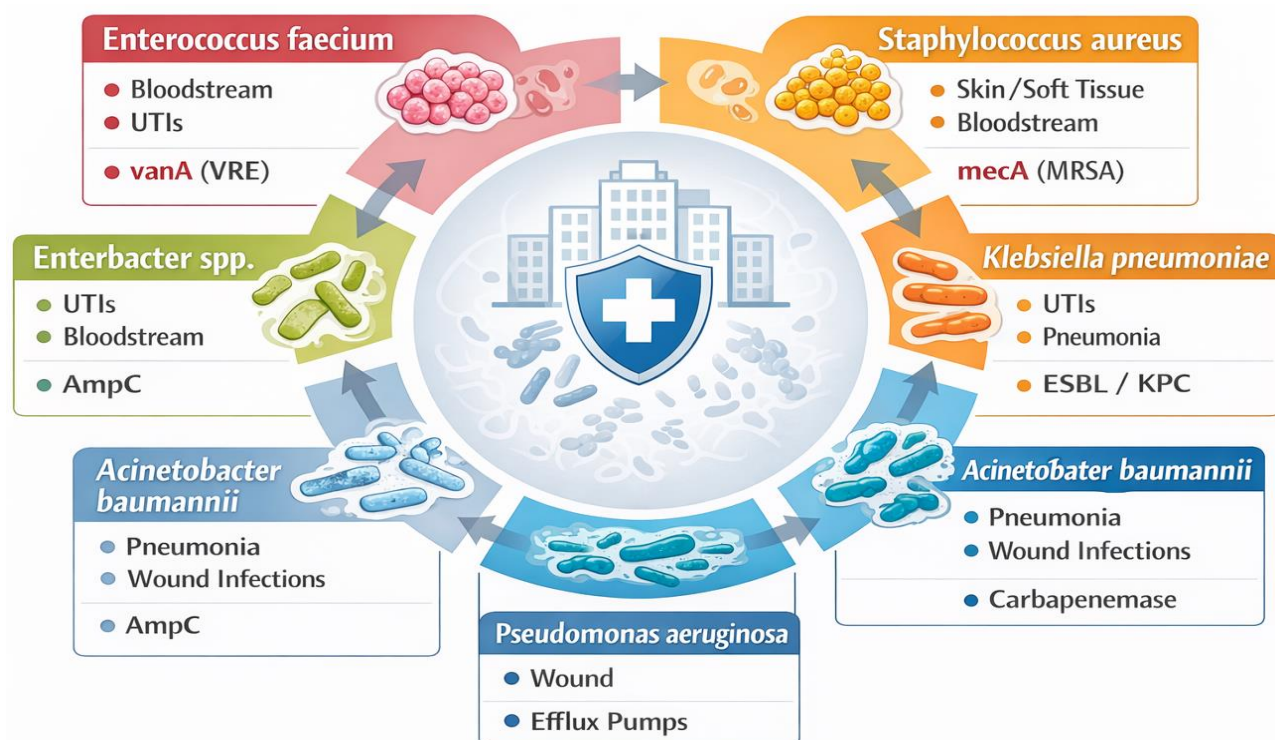
Table 2: Multidrug-Resistant (MDR) Detection Rates and Resistance Patterns by Clinical Specimen Type

Specimen Type	MDR Detection Rate (%)	Most Common MDR Pathogens	Notable Resistance Pattern
Urine	72%	<i>E. coli</i> , <i>K. pneumoniae</i>	ESBL, Fluoroquinolone resistance (Pan et al., 2025; Diop et al., 2025)
Blood	52.3%	<i>S. typhi</i> , MRSA, <i>Klebsiella</i>	XDR, 3-Generation Cephalosporin resistance (Ullah et al., 2025; Elmubark et al., 2025)
Wound/Pus	41.2%	<i>S. aureus</i> , <i>P. aeruginosa</i>	MRSA, Biofilm-associated resistance (Shrestha et al., 2024; Elmubark et al., 2025)
Sputum	-	<i>A. baumannii</i> , <i>P. aeruginosa</i>	DTR, Carbapenem resistance (Ullah et al., 2025; Genovese et al., 2025)
CSF	-	<i>Proteus</i> sp., <i>S. aureus</i>	High mortality risk (Elmubark et al., 2025)

The ESKAPE Pathogens: Biology, Mechanisms, and Clinical Impact

The ESKAPE pathogens represent a group of microorganisms that have developed sophisticated strategies to evade antibiotic action, making them the primary contributors to nosocomial morbidity and mortality (Ma et al., 2020). The major multidrug-resistant organisms responsible for healthcare-associated infections are collectively known as the ESKAPE pathogens, as illustrated in figure 2. These organisms possess diverse resistance mechanisms that enable them to evade the effects of multiple antibiotic classes.

Figure 2: The ESKAPE Pathogens and Their Clinical Significance



Enterococcus faecium: The Resilient Commensal

Enterococcus faecium has transitioned from a commensal of the human gut to a major hospital-acquired pathogen, characterized by intrinsic resistance to many beta-lactams and its ability to acquire high-level vancomycin resistance (VRE) (Huang et al., 2025). Between 2011 and 2014, *Enterococcus* species accounted for 14% of hospital-acquired infections in the United States, with *E. faecium* showing higher resistance rates than *E. faecalis* (Fialho, 2025). In Africa, *E. faecium* resistance is particularly high against tetracycline (47%), while in China, the prevalence of VRE_{fm} reached 2.2% in 2022. The genetic basis for vancomycin resistance typically involves the *vanA*, *vanB*, or *vanM* genes (Luo et al., 2024).

Staphylococcus aureus: The Versatile Pathogen

Staphylococcus aureus is a versatile pathogen capable of causing infections ranging from minor skin abscesses to life-threatening endocarditis. Methicillin resistance, mediated by the *mecA* gene which encodes an alternative penicillin-binding protein (PBP2a), is the hallmark of MDR *S. aureus* (Khasapane et al., 2025).

Klebsiella pneumoniae: The Carbapenemase Producer

Klebsiella pneumoniae is a leading cause of healthcare-associated pneumonia and is a major reservoir for highly mobile resistance genes. It is a prominent producer of ESBLs and carbapenemases, including *Klebsiella pneumoniae* carbapenemases (KPC), New Delhi metallo-beta-lactamases (NDM), and OXA-48-like enzymes (Ghanbarinasab et al., 2023). In Pakistan, ESBL production was observed in 83.3% of *Klebsiella* isolates, while carbapenem-resistant Enterobacteriaceae (CRE) rates reached 62.8% (Zhang et al., 2018).

Acinetobacter baumannii: The Hardest to Treat

Acinetobacter baumannii is often considered the most challenging ESKAPE pathogen to treat due to its rapid acquisition of resistance. It utilizes multiple mechanisms, including carbapenemase production, efflux pumps, and loss of porins (Mulani et al., 2019). In South Africa, over 80% of clinical isolates are MDR, and in Pakistan, 84%

are classified as difficult-to-treat resistance (DTR) strains (GBD 2021 Antimicrobial Resistance Collaborators, 2024).

Pseudomonas aeruginosa: The Efflux Specialist

Pseudomonas aeruginosa is notorious for its large genome and complex regulatory networks. It frequently employs RND-family efflux pumps and inducible AmpC beta-lactamases (Salleh et al., 2025). While it generally maintains higher susceptibility to colistin compared to *Acinetobacter*, resistance to carbapenems and fourth-generation cephalosporins is rising. In a five-year meta-analysis, the prevalence of MDR and XDR *P. aeruginosa* in Asia and Africa was 46.0% and 19.6%, respectively (Fong, 2023).

Enterobacter Species: The Inducible Resistor

Members of the *Enterobacter* genus are common causes of bacteremia in ICU settings. They are characterized by chromosomally encoded AmpC enzymes that can be induced by exposure to certain antibiotics, leading to clinical failure (Drozdzinsky et al., 2021). In African studies, *Enterobacter* spp. showed the highest resistance to amoxicillin-clavulanate (95.3%) (Anju et al., 2020).

Table 3: Resistance Profiles and Last-Resort Therapeutic Options for ESKAPE Pathogens

Pathogen	Primary Resistance Genes	Typical Phenotype	Preferred Therapy	Last-Resort
<i>E. faecium</i>	<i>vanA</i> , <i>vanB</i> , <i>vanM</i>	VRE, Linezolid-R (rare)	Linezolid, Daptomycin (Zhang et al., 2024; Pan et al., 2025)	
<i>S. aureus</i>	<i>mecA</i> , <i>blaZ</i>	MRSA	Vancomycin, Linezolid (Sah et al., 2023; Khasapane et al., 2025)	
<i>K. pneumoniae</i>	<i>blaCTX-M</i> , <i>blaKPC</i> , <i>blaNDM</i> , <i>blaOXA-48</i>	ESBL, CRE	Tigecycline, Colistin (Ghanbarinasab et al., 2023; Pan et al., 2025)	
<i>A. baumannii</i>	<i>blaOXA-23-like</i> , <i>blaNDM</i>	MDR, XDR, DTR	Colistin, Cefiderocol (Giordano et al., 2023; Sah et al., 2023)	
<i>P. aeruginosa</i>	<i>MexAB-OprM</i> (efflux), <i>blaVIM</i>	MDR, DTR	Colistin, Ceftolozane/tazobactam (Maleki et al., 2024)	
<i>Enterobacter</i> spp.	<i>AmpC</i> , <i>qnrS</i>	3rd Gen Ceph-resistant	Carbapenems, Aminoglycosides (Sah et al., 2023; Khasapane et al., 2025)	

Molecular Determinants and Mechanisms of Resistance

The adaptive evolution of MDR bacteria is driven by a sophisticated repertoire of molecular mechanisms (Fodor et al., 2020).

Enzymatic Inactivation and Modification

The production of beta-lactamases is the most prevalent mechanism of resistance against the beta-lactam class. ESBLs, predominantly of the CTX-M family, hydrolyze penicillins and third-generation cephalosporins but are inhibited by clavulanic acid (Akhtar et al., 2022). Carbapenemases represent a more severe threat. Molecular class B metallo-beta-lactamases (MBLs), such as NDM-1, require zinc ions and are not inhibited by common beta-lactamase inhibitors (Ahmed, 2021). Class D OXA-48 carbapenemases are notable for their ability to hydrolyze carbapenems while often

sparing extended-spectrum cephalosporins unless co-expressed with an ESBL (Hussain al., 2021).

Target Site Modification

Bacteria can also avoid antibiotic action by altering the molecular structure of the drug's target. This is seen in MRSA, where the *mecA* gene provides a penicillin-binding protein (PBP2a) with low affinity for beta-lactams (Otarigho et al., 2023). Similarly, vancomycin resistance in *Enterococcus* involves the modification of the cell wall precursor terminus to reduce vancomycin's binding affinity. Resistance to fluoroquinolones often arises from mutations in the *gyrA* and *parC* genes (Ambade et al., 2023).

Efflux Pumps and Reduced Permeability

The active expulsion of antibiotics is mediated by efflux pumps, particularly those of the RND family. These pumps can transport a wide range of structurally unrelated compounds, conferring an MDR phenotype (Colclough et al., 2020). Reduced membrane permeability, often through the loss of outer membrane porins, restricts antibiotic entry (Henderson et al., 2021).

Horizontal Gene Transfer and Plasmid Dissemination

The rapid spread of resistance is largely facilitated by horizontal gene transfer. Plasmids serve as major vectors for resistance genes, often carrying multiple genes that allow a single antibiotic to select for a strain resistant to many drug classes (Singh et al., 2025).

Table 4: Molecular Mechanisms of Resistance and Associated Antibiotic Classes

Mechanism Category	Molecular Examples	Targeted Antibiotic Classes	Primary Host Pathogens
Enzymatic Degradation	<i>bla</i> NDM-1, <i>bla</i> KPC-2, <i>bla</i> CTX-M-15	Beta-lactams, Carbapenems	<i>K. pneumoniae</i> , <i>E. coli</i> (Ghanbarinasab et al., 2023; Sah et al., 2023)
Target Modification	<i>mecA</i> , <i>vanA</i> , <i>gyrA</i> mutations	Methicillin, Vancomycin, Quinolones	<i>S. aureus</i> , <i>E. faecium</i> (Zhang et al., 2024; Sah et al., 2023)
Efflux Overexpression	<i>MexAB-OprM</i> , <i>AcrAB-TolC</i>	Macrolides, Tetracyclines, Quinolones	<i>P. aeruginosa</i> , <i>A. baumannii</i> (GBD 2021 Antimicrobial Resistance Collaborators, 2024; Maleki et al., 2024)
Target Protection	<i>qnrA</i> , <i>qnrB</i> , <i>qnrS</i>	Fluoroquinolones	<i>Enterobacter</i> spp., <i>E. coli</i> (Sah et al., 2023)
Altered Permeability	<i>OmpK35/36</i> loss	Carbapenems	<i>K. pneumoniae</i> (Ghanbarinasab et al., 2023)

The Role of Biofilms in Persistent and Recalcitrant Infections

A biofilm is a group of microorganisms encased in a self-produced extracellular matrix of polysaccharides, proteins, and extracellular DNA. It is estimated that up to 80% of human bacterial infections involve biofilms, particularly those associated with indwelling medical devices (Zafer et al., 2024).

Physical and Chemical Barriers to Antibiotic Action

The extracellular polymeric substance (EPS) matrix acts as a significant physical and chemical barrier. Infrared spectroscopy has demonstrated that the rate of transport for antibiotics like ciprofloxacin is significantly lower on colonized surfaces compared to sterile ones (Pipitò et al., 2025). The matrix also contains adsorption sites that can trap and accumulate antibiotic-degrading enzymes, such as beta-lactamases (Thomsen et al., 2023).

Metabolic Heterogeneity and "Persister" Cells

Within a biofilm, nutrient and oxygen gradients create diverse microenvironments, leading to stratified metabolic rates. Cells in the inner layers enter a state of dormancy, making them less sensitive to antibiotics that target actively dividing cells (Shamsrizi et al., 2020). Furthermore, biofilms harbor small subpopulations of "persister" cells that can survive extreme conditions only to repopulate the environment once the stress is removed (Sartorius et al., 2024).

Quorum Sensing and Genetic Exchange

Biofilm formation and maturation are regulated by quorum sensing, a cell-to-cell communication system that coordinates gene expression based on population density. Quorum sensing facilitates the upregulation of genes associated with drug resistance and efflux pumps (Mulani et al., 2019). Moreover, the close proximity of bacteria within a biofilm provides an ideal environment for the transfer of mobile genetic elements carrying resistance genes (Bale, 2021).

Table 5: Biofilm-Mediated Resistance Mechanisms Compared to Planktonic States

Biofilm Mechanism	Impact on Resistance	Comparison to Planktonic State	Clinical Significance
EPS Matrix Barrier	Reduced drug penetration	10 to 1,000-fold MIC increase (Yasir et al., 2024)	Failure of standard therapy (Yasir et al., 2024)
Metabolic Stratification	Tolerance in dormant cells	Inactive cells survive beta-lactams (Yasir et al., 2024)	Persistent/Chronic infections (Yasir et al., 2024)
Persister Cells	Spore-like survival state	100% susceptibility in planktonic (Yasir et al., 2024)	Recurrent infections after treatment (Yasir et al., 2024)
Quorum Sensing	Coordinated gene expression	Enhanced efflux and virulence (Maleki et al., 2024)	Targeted for "Quorum Quenching" (Yasir et al., 2024)
HGT Stability	Facilitated plasmid transfer	Higher rate of gene dissemination (Yasir et al., 2024)	Evolution of "Superbugs" (Yasir et al., 2024)

Socio-Economic and Environmental Drivers of Resistance

The emergence of MDR bacteria is rooted in human activities and environmental conditions (Ma et al., 2020).

Inappropriate Antibiotic Use and Stewardship Challenges

The misuse and overuse of antibiotics exert strong selective pressure that favors the survival of resistant strains. In LMICs, the situation is compounded by unregulated sales of antibiotics and lack of access to proper diagnostics (Stamkou et al., 2025). Healthcare professionals often face a dilemma in the emergency department, where the

need for rapid empirical treatment to prevent sepsis must be balanced against the risk of driving resistance (Ugochi Grace et al., 2025).

Environmental Reservoirs and "One Health" Perspective

AMR is increasingly detected in environmental reservoirs, including wastewater from hospitals and agricultural runoff. A "One Health" approach, which recognizes the interconnected health of people, animals, and the environment, is essential for understanding transmission cycles (Abebe et al., 2019).

Socio-Demographic Factors and Infrastructure

Overcrowded healthcare facilities, limited access to clean water, and inadequate sanitation infrastructure propagate the rapid transmission of resistant pathogens. Furthermore, global travel and trade act as conduits for the rapid transborder dissemination of resistant clones (Noreen et al., 2025).

Strategies for Containment and Antimicrobial Stewardship

Addressing the global health crisis of MDR bacteria requires effective antimicrobial stewardship (ASP), diagnostic innovation, and international collaboration (Pandey et al., 2021).

Diagnostic Innovations and Stewardship

The integration of rapid diagnostic tests (RDTs) is essential. Techniques such as polymerase chain reaction (PCR) and MALDI-TOF mass spectrometry can identify resistant strains much faster than traditional culture-based methods. "Diagnostic-driven de-escalation" leveraging biomarkers and rapid microbial identification allows for personalized therapy (World Health Organization, 2021).

Surveillance and Global Collaboration

Strengthening surveillance systems, particularly the WHO Global Antimicrobial Resistance and Use Surveillance System (GLASS), is critical. Standardized approaches for reporting resistance data enable reliable comparisons across different settings and countries (Pan et al., 2025).

Conclusion

Multidrug-resistant bacterial infections have evolved from a localized concern into a global public health emergency, with profound clinical, economic, and societal consequences. This review highlights alarmingly high MDR prevalence across clinical specimens particularly urine, blood, and wound samples and the dominance of ESKAPE pathogens that exhibit complex, multi-mechanism resistance profiles, including enzymatic inactivation, target alteration, efflux overexpression, reduced permeability, and robust biofilm formation. The situation is especially dire in LMICs of Africa and South Asia, where limited diagnostic capacity, unregulated antibiotic access, and inadequate infection control amplify transmission and mortality. The data clearly demonstrate that current empirical treatment regimens are increasingly ineffective, leading to prolonged hospital stays, higher treatment costs, and excess deaths. Without immediate intervention, the projected 10 million annual AMR-related deaths by 2050 will become reality. Effective containment demands a multifaceted strategy: rigorous antimicrobial stewardship programs, widespread adoption of rapid diagnostics and culture-guided therapy, strengthened national and global surveillance systems, and implementation of the One Health framework to address resistance at the human–animal–environment interface. Investment in new antibiotics, alternative therapies (e.g., phage therapy, anti-biofilm agents), and infection prevention infrastructure is equally critical. Only through coordinated, sustained global action can we slow the spread of MDR bacteria and safeguard the future of modern medicine.

References

- Bangladesh National Surveillance Team. (2024). *Antimicrobial resistance surveillance report: 2020-2024 clinical specimen analysis*. MDPI.
- Diop, M., Bassoum, O., Ndong, A., Wone, F., Tamouh, A. G., Ndoeye, M., Youbong, T., Daffé, S. M. M., Radji, R. O., Gueye, M. W., Lakhe, N. A., Fall, B., Ba, P. S., & Faye, A. (2025). Prevalence of multidrug-resistant bacteria in healthcare and community settings in West Africa: Systematic review and meta-analysis. *BMC Infectious Diseases*, 25(1), 292. <https://doi.org/10.1186/s12879-025-10562-w>
- Elmubark, A., Elmubark, M., & Abdallah, M. (2025). Prevalence and associated factors of multi-drug resistant bacteria among different clinical specimens at Wad Medani, Sudan: A four-year, cross-sectional study. *Scientific Reports*, 15(1), 714. <https://doi.org/10.1038/s41598-025-00714-y>
- GBD 2021 Antimicrobial Resistance Collaborators. (2024). Global burden of bacterial antimicrobial resistance 1990–2021: A systematic analysis with forecasted estimates to 2050. *The Lancet*, 404(10459), 1199–1226. ([https://doi.org/10.1016/S0140-6736\(24\)01867-1](https://doi.org/10.1016/S0140-6736(24)01867-1))
- Stamkou, D., Panteris, E., & Siasios, P. (2025). Antimicrobial resistance profiles of multidrug-resistant pathogens in a military hospital: A three-year study. *Cureus*, 17(8), Article e91297. <https://doi.org/10.7759/cureus.91297>
- Ghanbarinasab, F., Haeili, M., Ghanati, S. N., & Moghimi, M. (2023). High prevalence of OXA-48-like and NDM carbapenemases among carbapenem-resistant *Klebsiella pneumoniae* clinical isolates. *Journal of Medical Microbiology and Infectious Diseases*, 11(2), 78–85. <https://doi.org/10.52547/jmedmicrob.11.2.78>
- Pipitò, L., Rubino, R., Giammanco, A., Imburgia, C., Gaglio, A., Gagliardi, C., Giammanco, G. M., & Cascio, A. (2025). Antimicrobial resistance in ESKAPE pathogens: A retrospective epidemiological study at the University Hospital of Palermo, Italy. *Antibiotics*, 14(2), Article 186. <https://doi.org/10.3390/antibiotics14020186>
- Khasapane, N. G., Nkhebenyane, S. J., Lekota, K. E., Thekisoe, O., & Ramatla, T. (2025). An insight into the antibiotic resistance profiles of ESKAPE pathogens isolated from humans in Africa: A systematic review and meta-analysis. *BMC Infectious Diseases*, 25(1), 1730. <https://doi.org/10.1186/s12879-025-12109-5>
- Salleh, M. Z., Nik Zuraina, N. M. N., Deris, Z. Z., & Mohamed, Z. (2025). Current trends in the epidemiology of multidrug-resistant and beta-lactamase-producing *Pseudomonas aeruginosa* in Asia and Africa: A systematic review and meta-analysis. *PeerJ*, 13, Article e18986. <https://doi.org/10.7717/peerj.18986>
- Mudenda, S., Daka, V., & Matafwali, S. K. (2024). Diagnostic stewardship trends and antimicrobial resistance profiles of bacteria isolated in Zambia: A five-year retrospective study (2020–2024). *Antibiotics*, 13(8), 714. <https://doi.org/10.3390/antibiotics13080714>
- Noreen, N., Jamil, H., & Nuhrio, S. M. (2025). Uncovering the roadblocks: Challenges to implementing antimicrobial stewardship in Pakistan. *Link Medical Journal*, 3(1), 15–28. <https://doi.org/10.61919/zaedrv55>
- Pan, S., Agarwal, A., & Sharma, R. (2025). Prevalence and associated factors of multi-drug resistant bacteria among different clinical specimens in tertiary care hospital. *Journal of Neonatal Surgery*, 14(32s), 45–56.
- Sah, R. K., Bhattarai, A., & Khadka, P. (2023). Current antibiotic resistance profile of ESKAPE pathogens in a Nepalese hospital. *Cureus*, 15(5), e38421. <https://doi.org/10.7759/cureus.38421>
- Maharjan, S., Shrestha, S., & Shrestha, R. (2026). Prevalence and determinants of multi-drug resistance bacterial infection among burn patients in a tertiary care

- center in Nepal. *Infection and Drug Resistance*, 19, 1–13. <https://doi.org/10.2147/IDR.S559803>
- Ullah, A., Shabil, M., Abdulsamad, S. A., Jan, A., Naeem, A. A., Ullah, H., Khattak, M., & Ullah, Z. (2025). Prevalence of the antibiotic resistance of *Salmonella typhi* and *Salmonella paratyphi* in Pakistan: A systematic review and meta-analysis. *Open Forum Infectious Diseases*, 12(4), ofaf131. <https://doi.org/10.1093/ofid/ofaf131>
- World Health Organization. (2021). *Global antimicrobial resistance and use surveillance system (GLASS) report 2021*. <https://www.who.int/publications/i/item/9789240027336>
- Singh, B., Dahiya, M., Kumar, V., Ayyagari, A., Chaudhari, D. N., & Ahire, J. J. (2025). Biofilm and antimicrobial resistance: Mechanisms, implications, and emerging solutions. *Microbiology Research*, 16(8), Article 183. <https://doi.org/10.3390/microbiolres16080183>
- Luo, Q., Lu, P., Chen, Y., Shen, P., Zheng, B., Ji, J., Ying, C., Liu, Z., & Xiao, Y. (2024). ESKAPE in China: Epidemiology and characteristics of antibiotic resistance. *Emerging Microbes & Infections*, 13(1), Article 2317915. <https://doi.org/10.1080/22221751.2024.2317915>
- Gajic, I., Tomic, N., Lukovic, B., Jovicevic, M., Kekic, D., Petrovic, M., ... & Opavski, N. (2025). A comprehensive overview of antibacterial agents for combating Multidrug-Resistant bacteria: the current landscape, development, future opportunities, and challenges. *Antibiotics*, 14(3), 221.
- Karnwal, A., Jassim, A. Y., Mohammed, A. A., Al-Tawaha, A. R. M. S., Selvaraj, M., & Malik, T. (2025). Addressing the global challenge of bacterial drug resistance: insights, strategies, and future directions. *Frontiers in Microbiology*, 16, 1517772.
- Kashef Hamadani, B. H. (2023). The global geospatial distribution of antimicrobial resistance in selected gram-negative bacterial infections (Doctoral dissertation, University of Oxford).
- Sartorius, B., Gray, A. P., Weaver, N. D., Aguilar, G. R., Swetschinski, L. R., Ikuta, K. S., ... & Naghavi, M. (2024). The burden of bacterial antimicrobial resistance in the WHO African region in 2019: a cross-country systematic analysis. *The Lancet Global Health*, 12(2), e201-e216.
- Osen, G., Kapoor, G., Kalanxhi, E., Ouassa, T., Shumba, E., Brar, S., ... & MAAP Study Group. (2025). Antimicrobial resistance in Africa: A retrospective analysis of data from 14 countries, 2016–2019. *PLoS medicine*, 22(6), e1004638.
- Shamsrizi, P., Gladstone, B. P., Carrara, E., Luise, D., Cona, A., Bovo, C., & Tacconelli, E. (2020). Variation of effect estimates in the analysis of mortality and length of hospital stay in patients with infections caused by bacteria-producing extended-spectrum beta-lactamases: a systematic review and meta-analysis. *BMJ open*, 10(1), e030266.
- da Silva, K. E., Tanmoy, A. M., Pragasam, A. K., Iqbal, J., Sajib, M. S. I., Mutreja, A., ... & Andrews, J. R. (2022). The international and intercontinental spread and expansion of antimicrobial-resistant *Salmonella Typhi*: a genomic epidemiology study. *The Lancet Microbe*, 3(8), e567-e577.
- Mirha, H. T., Ali, S. H., Aamar, H., Sadiq, M., Tharwani, Z. H., Habib, Z., & Malikzai, A. (2024). The impact of antibiotic resistance on the rampant spread of infectious diseases in Pakistan: Insights from a narrative review. *Health science reports*, 7(4), e2050.
- Pandey, R., Mishra, S. K., & Shrestha, A. (2021). Characterisation of ESKAPE pathogens with special reference to multidrug resistance and biofilm production in a Nepalese hospital. *Infection and Drug Resistance*, 2201-2212.

- Wise, M. G., Karlowsky, J. A., Mohamed, N., Hermesen, E. D., Kamat, S., Townsend, A., ... & Sahm, D. F. (2024). Global trends in carbapenem-and difficult-to-treat-resistance among World Health Organization priority bacterial pathogens: ATLAS surveillance program 2018–2022. *Journal of global antimicrobial resistance*, 37, 168-175.
- Thomsen, J., Abdulrazzaq, N. M., UAE AMR Surveillance Consortium, Everett, D. B., Menezes, G. A., Senok, A., & Ayoub Moubareck, C. (2023). Carbapenem resistant Enterobacterales in the United Arab Emirates: a retrospective analysis from 2010 to 2021. *Frontiers in Public Health*, 11, 1244482.
- Abebe, M., Tadesse, S., Meseret, G., & Derby, A. (2019). Type of bacterial isolates and antimicrobial resistance profile from different clinical samples at a Referral Hospital, Northwest Ethiopia: five years data analysis. *BMC research notes*, 12(1), 568.
- Hoque, M., Mesbah, A., & Bala, A. S. (2024). The prevalence of multi-drug-resistant staphylococcus aureus and klebsiella pneumoniae in cooked meat samples collected from hospital cafeterias in the Dhaka city corporation area (Doctoral dissertation, Brac University).
- Bale, M. I. (2021). Molecular Characterization of Methicillin-Resistant Staphylococcus aureus (MRSA) Isolated from Blood Samples of Patients Attending Selected Hospitals in Ilorin, Nigeria (Doctoral dissertation, Kwara State University (Nigeria)).
- Ugochi Grace, O., Michael Ikechukwu, N., & Ogechi Innocentia, N. (2025). ANTIBIOTIC RESISTANT PROFILE AND MOLECULAR DETECTION OF RESISTANCE GENES IN MULTI-DRUG-RESISTANT Staphylococcus aureus ISOLATED FROM RETAIL MEAT IN OWERRI. *International Nexus Multidisciplinary Research Journal*, 2(1), 97-122.
- Ma, Y. X., Wang, C. Y., Li, Y. Y., Li, J., Wan, Q. Q., Chen, J. H., ... & Niu, L. N. (2020). Considerations and caveats in combating ESKAPE pathogens against nosocomial infections. *Advanced Science*, 7(1), 1901872.
- Huang, C., Moradi, S., Sholeh, M., Tabaei, F. M., Lai, T., Tan, B., ... & Azizian, K. (2025). Global trends in antimicrobial resistance of Enterococcus faecium: a systematic review and meta-analysis of clinical isolates. *Frontiers in Pharmacology*, 16, 1505674.
- Fialho, N. M. D. S. (2025). Antimicrobial drug resistance of Enterococcus spp. from humans, animals and the environment: a One Health approach.
- Zhang, Y., Wang, Q., Yin, Y., Chen, H., Jin, L., Gu, B., ... & Wang, H. (2018). Epidemiology of carbapenem-resistant Enterobacteriaceae infections: report from the China CRE network. *Antimicrobial agents and chemotherapy*, 62(2), 10-1128.
- Mulani, M. S., Kamble, E. E., Kumkar, S. N., Tawre, M. S., & Pardesi, K. R. (2019). Emerging strategies to combat ESKAPE pathogens in the era of antimicrobial resistance: a review. *Frontiers in microbiology*, 10, 539.
- Fong, I. W. (2023). New Cephalosporins: fifth and sixth generations. In *New Antimicrobials: For the Present and the Future* (pp. 25-38). Cham: Springer International Publishing.
- Drozdinsky, G., Neuberger, A., Rakedzon, S., Nelgas, O., Cohen, Y., Rudich, N., ... & Yahav, D. (2021). Treatment of bacteremia caused by Enterobacter spp.: should the potential for ampC induction dictate therapy? A retrospective study. *Microbial Drug Resistance*, 27(3), 410-414.
- Anju, V. T., Siddhardha, B., & Dyavaiah, M. (2020). Enterobacter infections and antimicrobial drug resistance. In *Model organisms for microbial pathogenesis, biofilm formation and antimicrobial drug discovery* (pp. 175-194). Singapore: Springer Singapore.

- Fodor, A., Abate, B. A., Deák, P., Fodor, L., Gyenge, E., Klein, M. G., ... & Makrai, L. (2020). Multidrug resistance (MDR) and collateral sensitivity in bacteria, with special attention to genetic and evolutionary aspects and to the perspectives of antimicrobial peptides—a review. *Pathogens*, 9(7), 522.
- Akhtar, A., Fatima, N., & Khan, H. M. (2022). Beta-lactamases and their classification: an overview. *Beta-Lactam Resistance in Gram-Negative Bacteria: Threats and Challenges*, 25-33.
- Ahmed, D. A. (2021). BETA-LACTAMASES ENZYMES: MECHANISM AND CLASSIFICATION. *Biochemical & Cellular Archives*, 21.
- Hussain, H. I., Aqib, A. I., Seleem, M. N., Shabbir, M. A., Hao, H., Iqbal, Z., ... & Li, K. (2021). Genetic basis of molecular mechanisms in β -lactam resistant gram-negative bacteria. *Microbial pathogenesis*, 158, 105040.
- Otarigho, B., & Falade, M. O. (2023). Computational screening of approved drugs for inhibition of the antibiotic resistance gene *mecA* in methicillin-resistant *Staphylococcus aureus* (MRSA) strains. *BioTech*, 12(2), 25.
- Ambade, S. S., Gupta, V. K., Bhole, R. P., Khedekar, P. B., & Chikhale, R. V. (2023). A review on five and six-membered heterocyclic compounds targeting the penicillin-binding protein 2 (PBP2A) of methicillin-resistant *Staphylococcus aureus* (MRSA). *Molecules*, 28(20), 7008.
- Colclough, A. L., Alav, I., Whittle, E. E., Pugh, H. L., Darby, E. M., Legood, S. W., ... & Blair, J. M. (2020). RND efflux pumps in Gram-negative bacteria; regulation, structure and role in antibiotic resistance. *Future microbiology*, 15(2), 143-157.
- Henderson, P. J., Maher, C., Elbourne, L. D., Eijkelkamp, B. A., Paulsen, I. T., & Hassan, K. A. (2021). Physiological functions of bacterial “multidrug” efflux pumps. *Chemical reviews*, 121(9), 5417-5478.
- Zafer, M. M., Mohamed, G. A., Ibrahim, S. R., Ghosh, S., Bornman, C., & Elfaky, M. A. (2024). Biofilm-mediated infections by multidrug-resistant microbes: a comprehensive exploration and forward perspectives. *Archives of Microbiology*, 206(3), 101.