

## Molecular Mechanisms Linking Dietary Sugar to Metabolic Disorders, Inflammation, and Premature Aging

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### Abstract

Over consumption of dietary sugars, especially high fructose and sucrose, are now identified as one of the important factors contributing towards metabolic disturbance, chronic inflammation and accelerated aging process. Regular consumption of foods with high sugar content and sugar sweetened beverages is associated with obesity, insulin resistance, poor glucose tolerance, type 2 diabetes mellitus, non-alcoholic fatty liver disease (NAFLD) and cardiovascular disease. The results of molecular research and studies on populations suggest that chronic high sugar feeding can cause metabolic dysfunction in several linked ways, such as poor insulin signaling, greater lipid production in the liver, more oxidative damage and less efficient mitochondria. Unlike glucose metabolism, fructose metabolism stimulates triglyceride deposition, uric acid formation and metabolic irregularities because it is largely metabolized in the liver

without passing through critical steps in glycolysis. In addition, too much sugar activates inflammatory pathways such as NF- $\kappa$ B, MAPK pathways and the activation of the NLRP3 inflammasome, all of which play a role in chronic inflammatory reactions. Changes in gut microbes, combined with impaired gut barrier function may also amplify systemic inflammation and metabolic dysfunction. Moreover, chronic

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hyperglycemia is associated with increased generation of reactive oxygen species (ROS) and advanced glycation end products (AGEs) causing oxidative stress, DNA damage, dysfunction and senescence-related processes that increase the biological ageing process. High intake of sugar has also been shown to alter DNA methylation and histone modification, and it is emerging that these epigenetic changes may modulate the expression of genes related to metabolic and inflammatory pathways. Together, these findings indicate that chronic consumption of high-sugar diets has a wide-ranging molecular and physiological impact that negatively affects metabolic health and aging and suggest a need for nutritional intervention and prevention strategies to promote improved long-term health outcomes and healthy aging.

### **Introduction:**

Metabolic disorders continue to increase, as of today, so much so that it has become immensely worse than it was a few decades ago. This pattern may partly be attributed to the modern eating habits, as eating many industrially processed foods is an important contributor to the consumption of refined carbohydrates and added sugars (Malik *et al.*, 2010; Stanhope, 2016). Sugars are the main energy substrates and precursors for biosynthetic pathways, and if consumed at excess levels, likely to have more than just an excess caloric intake as a problem, such as a rapid rate of metabolism or even ageing of the cells. There is a clear link between the increase of sugar consumption and a higher risk of developing type 2 diabetes mellitus, obesity and decreased insulin sensitivity, as revealed by many studies. In addition, the metabolic derangements are commonly associated with cardiovascular diseases as well as non-alcoholic fatty liver disease (NAFLD).

Current research indicates that these conditions are not independent phenomena but are connected through common metabolic disturbances and overlapping molecular mechanisms. In these ways, but how exactly it all leads to aging faster, still cannot be well understood (Stanhope, 2016; Younossi *et al.*, 2016). Overall, changes in diet represent a major contributing factor that is likely to be supporting much of this. These associations are supported by epidemiological evidence. However, now it appears to be more concerned with an attempt to know the biochemical side of it, it is not only excess caloric intake alone (Y. Huang *et al.*, 2023; Ma *et al.*, 2022).

Sugars are also important components in the formation of molecules that are required for the growth of cell and cellular functions. Studies suggest the impact of sugars on metabolism is simply dependent on the type of sugar and how quickly it is absorbed, along with all the factors that come into play once it is in the body (Hannou *et al.*, 2018; Tappy and Lê, 2010). Different sugars exhibit distinct metabolic behaviors and physiological effects. In today's diet, there is a lot of processed food that contains sucrose or corn syrup with high fructose, and is consumed frequently. This leads to rapid increases in blood sugar and an imbalance in the hormones that are supposed to regulate the body. This dietary pattern is likely to disrupt the body's natural processes of metabolism, particularly with insulin and the processing of fats. This leads to a less flexible body in terms of energy sources, and glucose regulation becomes abnormal (Goodpaster and Sparks, 2017). These findings suggest a mechanistic association, although some mechanisms remain incompletely understood. (Alam *et al.*, 2022; C.) Huang *et al.*, 2023. New researches are emerged to show, glucose and fructose, despite having similar calorie values, have different metabolic pathways and regulatory mechanisms, which may account for the differences in their effects on metabolism.

Besides energy metabolism, cellular stress and inflammatory pathways have also been found to be activated with excessive sugar consumption. Chronic, mild systemic inflammation, also known as metaflammation, can be exacerbated by high dietary sugar intake. The development of metabolic syndrome and other chronic illnesses is likely to be greatly exacerbated by this situation. Mechanistically, mitochondrial

stress, elevated development of reactive oxygen species (ROS), and enhanced signals that causes inflammation are collectively connect nutritional excess with disturbances in immune and endocrine function (Chen *et al.*, 2015; Ma *et al.*, 2022). Growing attention has also focused on the way fructose is processed in the liver due to its potential metabolic implications relatively uncontrolled influx into glycolytic pathways, resulting in the production of uric acid, de novo lipogenesis, and metabolic stress that can further activate the inflammatory pathway and lipid accumulation (Baharuddin, 2025; Shi *et al.*, 2021).

Moreover, the metabolic and inflammatory responses, the effects of dietary sugars on biological processes of aging might be through non-enzymatic glycation, oxidative stress, and nutrient-sensing pathway dysregulation mechanisms. The build-up of the advanced glycosylation end-products (AGEs) is a significant biochemical process that connects the excess exposure to sugars to the dysfunction of the cell since AGEs can modify proteins and nucleic acids, influence cellular shape, and stimulate inflammatory reactions by the involvement of receptors (Aragno and Mastrocola, 2017; Semchyshyn, 2024; Uribarri *et al.*, 2010; Vlassara and Uribarri, 2014). These molecular processes intersect well-known age-associated phenotypes, including mitochondrial dysfunction, genomic instability, and defective cell interactions (López-Otín *et al.*, 2013), suggesting that excess sugar exposure can hasten the age-related deterioration of physiological performance (Franceschi *et al.*, 2018).

Considering the complexity of interactions between nutrient metabolism, immunity, and the aging of cells, the comprehensive knowledge of the biochemical characteristics of dietary sugars and molecular mechanisms of their action is urgently needed (López-Otín *et al.*, 2013; Franceschi *et al.*, 2018). In order to better understand the mechanisms underlying the relationship between dietary sugar consumption and the development of metabolic disorders, chronic inflammatory conditions, and aging-related processes, one has a unique opportunity to integrate data from the fields of biochemistry, molecular biology, and system-level research (Johnson *et al.*, 2013; Hannou *et al.*, 2018; Wang *et al.*, 2025). That's why, this article aims to examine the biochemical characteristics of dietary sugars, explore their metabolic processing within the body, and discuss the molecular mechanisms through which excessive sugar intake influences biological functions and health outcomes (Tappy and Lê, 2010; Lustig *et al.*, 2012; Stanhope, 2016; Malik *et al.*, 2010).

### **Dietary Sugars: Biochemical Characteristics and Metabolic Fate**

Dietary sugars are a diversified family of carbohydrates differing in biochemical properties, control, and impacts (Hannou *et al.*, 2018; Tappy and Lê, 2010). The absorption, metabolism, and usage of the diet sugars in the body are determined by the biochemical properties of sugars (Ferraris, 2001; Rippe and Angelopoulos, 2016). Even though sugar in all forms is an energy-giving compound, molecular structure differences influence the enzymatic reactions, hormonal regulation, and signal transduction (Lustig *et al.*, 2012; Malik *et al.*, 2010). The modern-day nutritional environment of excessive refined sugar consumption has enhanced the exposure to fast-absorbable carbohydrates, which influence the metabolic homeostasis (Johnson *et al.*, 2013; Popkin and Hawkes, 2016). The biochemical properties and metabolism of dietary sugars are significant to comprehend their impacts on the metabolic disorders, inflammation, and aging (Alam *et al.*, 2022; Hannou *et al.*, 2018).

### **Types of Dietary Sugars**

Dietary sugars may primarily be defined by the chemical structure as monosaccharides and disaccharides. The different substances have varying absorption patterns and metabolism in the two categories in which they exist (Tappy and Lê, 2010). Monosaccharides, including fructose, glucose, and galactose, are the simplest known carbohydrate structures. They are completely absorbed through the intestinal

lining of the body and do not require digestion. Glucose is the main sugar circulating in the blood that is regulated by the hormones through hormonal processes of the endocrine system (Ferraris, 2001). Fructose is broken down by the liver into different sets of enzymes. Sucrose, along with lactose and maltose are disaccharide molecule that is made up of two units of monosaccharides and bound together by glycosidic linkages. These need enzymes in the body to be broken down into forms that can be absorbed in the body. Sugars may also be classified as intrinsic sugars, naturally present within whole foods, or added sugars, which are incorporated during food processing. Individuals who consume added sugars have increased metabolic health issues as these sugars are absorbed into the blood quickly, and they easily overconsume such foods (Huang *et al.*, 2023; O'Connor *et al.*, 2018). Other methods of classifying sugars include the glycemic index and glycemic load, which provide the rate at which a certain sugar raises the blood glucose levels. Sugar sources with a high glycemic index result in rapid rises of glucose and insulin levels, whereas lower ones might exert more progressive action. Repeated postprandial glucose and insulin spikes may take part in the resistance in insulin and metabolic dysfunction of metabolism (Ludwig, 2002). The body reacts to different types of carbohydrates in the diet because their chemical components change how the body works and can lead to sickness. The classification, metabolic characteristics, and regulatory features of common dietary sugars are discussed in Table 1.

**Table 1. Classification, Metabolic features , and Regulatory Features of Common Dietary Sugars**

<b>Sugar Type</b>	<b>Classification</b>	<b>Primary Sources</b>	<b>Main Metabolic Site</b>	<b>Regulatory Control</b>	<b>Glycemic Response</b>	<b>References</b>
Glucose	Monosaccharide	Fruits, starch digestion products	Multiple tissues (muscle, brain, liver)	Strong insulin-mediated regulation	High glycemic response	Petersen and Shulman (2018); Herman and Birnbaum (2021)
Fructose	Monosaccharide	Fruits, honey, sucrose, high-fructose corn syrup (HFCS)	Primarily liver	Minimal direct insulin regulation; rapid hepatic metabolism	Low glycemic index but promotes lipogenesis	Hannou <i>et al.</i> (2018); Softic <i>et al.</i> (2020); Lanaspá <i>et al.</i> (2020)
Galactose	Monosaccharide	Dairy products (from lactose hydrolysis)	Liver (converted to glucose intermediates)	Moderate metabolic regulation	Moderate glycemic response	Stanhope (2020); Huang <i>et al.</i> (2023)

Sucrose	Disaccharide (Glucose + Fructose)	Table sugar, + processed foods	Hydrolyzed in intestine → glucose + fructose	Mixed regulation via component + monosaccharides	High glycemic response	Stanhope (2020); Softic <i>et al.</i> (2020)
Lactose	Disaccharide (Glucose + Galactose)	Milk and + dairy products	Hydrolyzed by lactase before absorption	Enzyme-dependent digestion	Moderate glycemic response	Huang <i>et al.</i> (2023); Petersen and Shulman (2018)

### Glucose Metabolism and Regulation

The glucose metabolism is one of the most highly regulated biochemical processes in human physiology, as glucose is a prime energy source (Murray *et al.*, 2018; Petersen and Shulman, 2018). Following consumption, glucose is absorbed in the small intestine by sodium-glucose cotransporter 1 (SGLT1) and enters the bloodstream, where it causes pancreatic  $\beta$ -cells to secrete insulin. Through the insulin-dependent glucose transporter GLUT4, insulin, the most crucial hormone for control, encourages the use of sugar in peripheral tissues, particularly skeletal muscle and adipose tissue (Herman and Birnbaum, 2021; Petersen and Shulman, 2018). When glucose is taken into cells, it goes through the process of glycolysis to produce pyruvate that may be used in mitochondrial oxidative phosphorylation to make ATP or may be utilized in anabolic pathways such as glycogen production and lipid synthesis.

The metabolism of glucose must be maintained relatively carefully, and otherwise the balance in energy turns out of shape and might facilitate the progression of metabolic disorders (Petersen and Shulman, 2018). This regulation is essential for maintaining metabolic homeostasis. These feedback loops consist of the insulin and glucagon loops, some of the nutrient sensory pathways, all of which are linked to one another to coordinate the use of glucose with body requirements at a given moment (Saltiel and Kahn, 2001; Rorsman and Ashcroft, 2018). However, the prolonged insulin signaling in the long run when glucose is continually elevated, such as when eating excess sugar causes it to begin to malfunction and may result in resistance.

At the molecular level, chronic hyperglycemia increases reactive oxygen species production, activates inflammatory transcription factors, and promotes non-enzymatic glycation (Brownlee, 2001; Giacco and Brownlee, 2010). All those are attributable to oxidative stress and actual cellular damage; it is proposed that glucose is regulated much more strongly than other sugars (Chen *et al.*, 2015; Ma *et al.*, 2022).

### Hepatic Metabolism of Fructose

Fructose is processed in a different way than glucose, primarily in the liver, and it bypasses some of the normal controls that govern the metabolism of carbohydrates (Tappy and L e, 2010; Stanhope, 2016). This may represent a major contributor to its metabolic effects. After intestinal absorption via GLUT5 transporters, fructose is rapidly used up by liver cells and convert into the fructose-1-phosphate by fructokinase (Ferraris, 2001). This process occurs largely independently of insulin and bypasses phosphofructokinase, a major regulatory step in glycolysis. This lack of regulation may promote the appearance of large quantities of fructose that would raise the number of building blocks to make fat, such as triglycerides, and that may accumulate in the liver or disrupt lipid levels generally (Hannou *et al.*, 2018; Shi *et al.*, 2021). Fructose does increase uric acid and ATP depletion, too, and the precise

mechanisms remain unclear, but it is likely related to the stress of oxygen and inflammation of metabolism (Hannou *et al.*, 2018; Johnson *et al.*, 2013).

These outcomes are more pronounced in the current diets that contain all the added sugar, including high-fructose corn syrup. Experiments indicate that fructose chronically impairs mitochondrial function, accumulates lipids in liver cells, and initiates inflammatory pathways that promote the progression of metabolic disease (Baharuddin, 2025; Lustig *et al.*, 2012). The less tightly regulated hepatic metabolism of fructose may explain its association with these issues, although some mechanisms remain unclear (Malik *et al.*, 2010; Popkin and Hawkes, 2016).

### **Comparative Molecular Effects of Glucose and Fructose**

Both glucose and fructose contain similar calories, yet the manner in which these substances are processed in the body occurs in a somewhat different manner at the molecular level. However, glucose is tightly regulated with the aid of insulin, making sure that glucose enters the cells and is converted to energy or stored. However, with fructose, there are several regulatory steps that are ignored, and the fructose is processed mainly in the liver. The outcome of this is that they have huge variations in their impact on fats, hormones and inflammation. For example, fructose is more likely than glucose to promote the production of fat in the liver, thereby increasing triglyceride levels and the risk of triglyceride accumulation in the liver. These changes may take part to the development of fatty liver disease and broader metabolic disturbances (Hannou *et al.*, 2018). This effect is mainly attributed to the production of metabolic intermediates during the catabolism of fructose that are used to de novo lipogenesis and then the storage in the liver. Fructose appears to exert a larger effect on lipid metabolism than glucose. Both sugars can induce oxidative stress and glycation reactions when consumed in excess. Nevertheless, fructose appears more reactive in such non-enzymatic mechanisms, accelerating the final products of the advanced glycation process that is more harmful to cells. These mechanisms are complex; however, this further exacerbates cellular damage (Shi *et al.*, 2021).

Furthermore, there are the nutrient sensing pathways, such as the AMPK and mTOR, which each sugar activates differently. These pathways may influence mitochondrial function and metabolic flexibility. It has been proposed that the reason why not all sugars are equally healthy, particularly with regard to doing something to the inflammation or aging process. In general, it is important to distinguish between glucose and fructose when considering diets and metabolism. It highlights the fact that these molecular pathways are not the same. Differences are notable in their impact on all the lipids to the signaling (Aragno and Mastrocola, 2017; Semchyshyn, 2024). The key molecular differences between glucose and fructose metabolism are mentioned in Table 2.

**Table 2. Comparative Molecular and Metabolic Characteristics of Glucose and Fructose Metabolism**

<b>Feature</b>	<b>Glucose</b>	<b>Fructose</b>	<b>Key References</b>
<b>Primary Absorption Transporter</b>	<i>Sodium–Glucose Linked Transporter 1</i> and GLUT transporters	Primarily GLUT5 transporter	Softic <i>et al.</i> (2020); Hannou <i>et al.</i> (2018)
<b>Major Metabolic Site</b>	Multiple tissues including muscle, brain, and liver	Predominantly metabolized in the liver	Hannou <i>et al.</i> (2018)
<b>Insulin</b>	Strong stimulation of	Minimal direct	Herman and

<b>Response</b>	insulin secretion	stimulation of insulin secretion	Birnbaum (2021)
<b>Entry into Glycolysis</b>	Regulated via phosphofruktokinase (PFK) control	Bypasses the major glycolytic regulatory step	Softic <i>et al.</i> (2020)
<b>Lipogenic Potential</b>	Moderate lipogenic activity	High lipogenic potential through enhanced <i>de novo</i> lipogenesis	Lanaspa <i>et al.</i> (2020)
<b>Uric Acid Production</b>	Minimal production	Increased uric acid generation	Lanaspa <i>et al.</i> (2020)
<b>Oxidative Stress Potential</b>	Moderate under excessive intake	Higher oxidative stress due to rapid hepatic metabolism	Rehman and Akash (2020)
<b>Association with Metabolic Disease</b>	Chronic excess linked to insulin resistance	Strong correlation with metabolic dysregulation, dyslipidemia, and non-alcoholic fatty liver disease	Stanhope (2020); Huang <i>et al.</i> (2023)

### **Molecular Mechanisms Underlying Sugar-Induced Metabolic Disorders Insulin Resistance and Signal Transduction Defects.**

Insulin resistance caused by an imbalance of molecular pathways involved in the balance of glucose homeostasis in the liver and peripheral tissues, including skeletal muscle and adipose tissue, has been linked to excessive consumption of dietary carbohydrates, particularly fructose (Basciano *et al.*, 2005). In contrast to glucose, fructose metabolism skips a number of significant regulatory steps in glycolysis and can be quickly metabolized into lipogenic intermediates. This metabolic pathway leads to increased accumulation of lipid intermediates like diacylglycerols (DAG), leading to activation of particular isoforms of protein kinase C (PKC). PKC enzyme activation results in serine phosphorylation of insulin receptor substrate proteins, which reduces insulin-dependent glucose absorption and the effectiveness of downstream PI3K/Akt signaling (Chen *et al.*, 2015; Herman and Birnbaum, 2021).

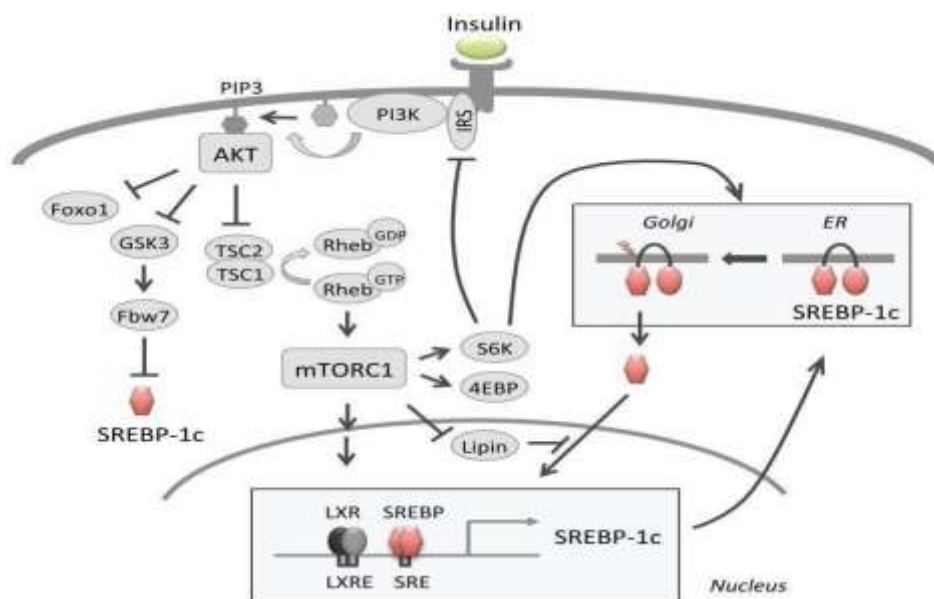
Fructose, further to its direct effect on signaling of insulin, also has lipotoxicity and keeps the chronic inflammatory state. Rutledge and Adeli (2007) have shown that accumulated lipid intermediates activate stress-sensitive signaling cycles like c-Jun N-terminal kinase (JNK) and nuclear factor-kappa B (NF-κB), which in turn decrease insulin responsiveness. Fructose intake has also been linked to increased expression of lipogenic regulators, such as sterol regulatory element-binding protein-1c (SREBP-1c) and carbohydrate-responsive element-binding protein (ChREBP), which increases the synthesis of hepatic triglycerides (HTG). This progressive lipid deposition into the liver eventually compounds the insulin resistance, which leads to ongoing metabolic abnormality.

Metabolism of fructose also causes impairment of cellular energy regulation and increases oxidative stress. Some of the intracellular ATP is used up by the increased production of uric acid and reactive oxygen species (ROS) brought on by the quick phosphorylation of fructose. These oxidative molecules along with the inflammatory mediators further worsen insulin sensitivity. (Sousa *et al.*, 2023) Furthermore, too much lipid in cells decreases the capacity to shift fibers between glucose and fat use, adds to the work of the mitochondria, and decreases metabolic flexibility (Murphy *et al.*, 2025). Experimental and clinical researches have shown that fructose intake is linked with a reduction in Akt phosphorylation in liver, skeletal muscle and adipose

tissue. Fasting insulin levels are increased and glucose tolerance becomes impaired with these changes, even with normal calorie intake, suggesting a direct inhibitory effect of fructose on insulin signaling pathways (Herman and Birnbaum, 2021). In conclusion, a number of interrelated mechanisms, including the effects on insulin intracellular signaling, lipid buildup in the liver and peripheral tissues, lipotoxicity, chronic inflammation, oxidative stress, uric acid production, and mitochondrial dysfunction, are associated with insulin resistance in rats fed fructose. The interplay of these processes can be a mechanistic explanation of the close relationship between high consumption of added sugars, mainly fructose, with type 2 diabetes mellitus, non-alcoholic fatty liver disease (NAFLD), and metabolic syndrome.

The binding of insulin to its cell surface receptor initiates an insulin signaling cascade, which sets up a series of internal cellular events that eventually result in the uptake of glucose. Phosphoinositide 3-kinase (PI3K) is triggered by receptors and changes phosphatidylinositol-4,5-bisphosphate (PIP<sub>2</sub>) into phosphatidylinositol-3,4,5-trisphosphate (PIP<sub>3</sub>) (Manning and Cantley, 2007). Phosphoinositide-dependent kinase-1 (PDK1) uses PIP<sub>3</sub> as a membrane docking site to activate protein kinase B (PKB/Akt). Numerous downstream targets, including forkhead box protein O1 (FoxO1), are regulated by activated Akt, which in turn controls the expression of metabolic genes including glucose-6-phosphatase (G6Pase) and phosphoenolpyruvate carboxykinase (PEPCK).

This is due to the fact that elevated quantities of free fatty acids in the blood can induce disruption by encouraging the serine phosphorylation of insulin receptor substrate (IRS) proteins, which reduces their normal tyrosine phosphorylation and impairs downstream insulin communication (Samuel and Shulman, 2012; Hotamisligil, 2017). The recruitment of IRS proteins and the spread of signals depend on the tyrosine phosphorylation of the insulin receptor. By activating PI3K, PDK1, and Akt, insulin signaling increases the absorption of glucose by the cell and encourages the translocation of glucose transporter 4 (GLUT4) to the plasma membrane. Important participants in this regulatory process are these signaling intermediates, namely PIP<sub>2</sub> and PIP<sub>3</sub>. Figure 1 shows the insulin signaling system and how metabolic stress disrupts it.



**Figure 1.** GLUT4 translocation to the cell membrane and cellular glucose uptake are ultimately facilitated by insulin binding to its receptor (IR), which under normal physiological conditions starts a signaling cascade involving insulin receptor

*substrate (IRS) proteins and activation of the PI3K/Akt pathway. Insulin resistance develops as a result of this signaling mechanism being disrupted under metabolic stress (Manning and Cantley, 2007; Samuel and Shulman, 2012). Serine phosphorylation of IRS proteins can be triggered by elevated levels of free fatty acids (FFAs) and pro-inflammatory signaling under metabolic stress. This can impede insulin signaling and increase insulin resistance.*

### **Lipotoxicity and De Novo Lipogenesis**

The process by which the liver turns extra carbohydrates into fat is known as hepatic de novo lipogenesis, and it is greatly accelerated by excessive dietary sugar, particularly fructose. Fructose is quickly transformed into triose phosphates and acetyl-CoA, bypassing major regulatory steps in glycolysis and providing substrates for fatty acid synthesis (Herman and Birnbaum, 2021). Hepatic steatosis, or fatty liver, and Lipotoxicity have been brought on by a similar process, which increases triglyceride synthesis in hepatocytes (Hannou *et al.*, 2018).

Fructose-driven lipogenesis is also mediated by the transcription factors SREBP-1c and ChREBP, which promote the production of enzymes such as acetyl-CoA carboxylase (ACC) and fatty acid synthase (Basciano *et al.*, 2005). By improving the liver's capacity to produce triglycerides, this raises hepatic fat buildup. In addition to hepatic injury, increased fat production causes VLDL export, or releases lipids into the blood and nearby tissues. Ceramides and diacylglycerols (DAG) are a couple of accumulating lipid intermediates that impede normal insulin signaling by activating PKC and stress kinases like NF- $\kappa$ B and JNK. This results in a chain reaction in that the lipid accumulation causes insulin resistance, which in turn promotes further lipid storage (Rutledge and Adeli, 2007). Experiments on humans and animals show that frequent consumption of sugar increases liver fat and increases the generation of VLDL, which causes lipids to accumulate in muscle and adipose tissue (Elliott *et al.*, 2002). By promoting peripheral tissue inflammation and metabolic dysfunction, this widespread lipotoxicity triggers the cycle of metabolic disease (MacDonald, 2016). Several ways exist by which insulin activates SREBP-1: Insulin induces transcription of SREBP-1c and induces proteolytic processing. It also induces nuclear localization of the processed protein and prevents proteosomal degradation of SREBP (Herman and Birnbaum, 2021).

### **Mitochondrial Dysfunction and Metabolic Inflexibility**

Excessive consumption of dietary sugar, particularly fructose, could impose significant metabolic demands on liver cell mitochondria, skeletal muscle mitochondria, and adipose tissue mitochondria. In this context, mitochondrial function is impaired, and ROS production is increased, cellular energy production is decreased, and fatty acid catabolism is less efficient (Hannou *et al.*, 2018). Thus, this capacity to switch usage from carbohydrate to lipid is lost, leading to decreased metabolic flexibility, this is a sign of malfunctioning metabolism (Basciano *et al.*, 2005; Taskinen *et al.*, 2019).

Fructose-driven de novo lipogenesis further increases lipid deposition, placing additional stress on mitochondrial function. Because lipid intermediates like ceramides and diacylglycerol (DAG) interfere with insulin signaling pathways and increase oxidative damage, mitochondrial dysfunction is associated with insulin resistance (Rutledge and Adeli, 2007). Mitochondrial ROS induces stress-related kinases, such as NF- $\kappa$ B and JNK, that lead to additional insulin-sensitising defects. Chronic fructose exposure also disrupts electron transport, promotes lipid peroxidation, and decreases the efficiency of the electron transport chain, which leads to a decrease in ETP efficiency and cellular energy production (Elliott *et al.*, 2002; MacDonald, 2016). These mitochondrial impairments contribute to the accumulation of lipotoxic intermediates, establishing a self-perpetuating cycle of oxidative stress,

disrupted energy metabolism, and metabolic dysfunction. All things considered, sugar-induced mitochondrial dysfunction increases oxidative stress, decreases metabolic flexibility, aggravates insulin resistance and Lipotoxicity, and contributes significantly to the development of metabolic syndrome.

### Uric Acid and Metabolic Stress

An excess quantity of uric acid are developed during the hepatic metabolism of fructose. Fructokinase quickly converts fructose to fructose-6-phosphate, using up the intracellular ATP, which increases levels of AMP, which is later catabolised via purine catabolism to uric acid (Herman and Birnbaum, 2021; Hannou et al., 2018). Elevated levels of uric acid have been associated with the activation of the NF- $\kappa$ B and JNK pathways, which interfere with regular insulin signaling, cause metabolic dysregulation, and increase oxidative stress (Rutledge and Adeli, 2007). Additionally, uric acid levels impair vascular function by lowering the bioavailability of nitric oxide. This malfunction limits the use of glucose by peripheral tissues like skeletal muscle and adipose tissue, compromises endothelium integrity, and lessens insulin's capacity to control vascular function (Basciano et al., 2005). This culminates in a vicious cycle of oxidative stress, inflammation, impaired mitochondrial function, and insulin resistance, which further worsen each other (Taskinen et al., 2019)

Numerous metabolic disorders, including obesity, type 2 diabetes mellitus, non-alcoholic fatty liver disease (NAFLD), and elements of the metabolic syndrome (MetS), have been consistently linked to hyperuricemia, according to epidemiological research and clinical studies (MacDonald, 2016). The information that is currently available makes it abundantly evident that uric acid, which is created during the metabolism of fructose, is a major mediator in the relationship between high sugar consumption and the emergence of metabolic disease due to its effects on insulin sensitivity, inflammation, and antioxidants (Elliott et al., 2002). Table 3 provides a summary of the main mechanisms at play.

**Table 3. Comparison of Key Mechanisms Involved in Sugar-Induced Metabolic Dysfunction**

<b>Mechanism</b>	<b>Biological Process</b>	<b>Major Molecular Components and Pathways</b>	<b>Physiological Consequence</b>	<b>References</b>
<b>Insulin Resistance</b>	Accumulation of lipid-derived metabolites disrupts normal insulin signaling	Diacylglycerol (DAG), Protein Kinase C (PKC), Insulin Receptor Substrate (IRS) serine modification, PI3K/Akt signaling pathway, c-Jun N-terminal kinase (JNK), and Nuclear Factor-kappa B (NF- $\kappa$ B)	Decreased cellular glucose utilization	Basciano et al. (2005); Herman and Birnbaum (2021)
<b>De Novo Lipogenesis</b>	transformation of metabolites generated from fructose into precursors of fatty acids	Fatty Acid Synthase (FASN), Acetyl-CoA Carboxylase (ACC), Carbohydrate-	Hepatic fat accumulation and elevated VLDL secretion	Hannou et al. (2018)

		Responsive Element-Binding Protein (ChREBP), and Sterol Regulatory Element-Binding Protein-1c (SREBP-1c)		
<b>Mitochondrial Dysfunction</b>	Excess nutrient availability impairs mitochondrial performance	Electron Transport Chain (ETC) disruption, JNK activation, NF-κB signaling, and reactive oxygen species (ROS)	Reduced metabolic adaptability and impaired energy regulation	Taskinen et al. (2019)
<b>Uric Acid-Associated Stress</b>	ATP depletion leading to enhanced purine degradation	Adenosine Monophosphate (AMP) catabolism, elevated uric acid production (hyperuricemia)	Endothelial impairment and metabolic stress	Elliott et al. (2002)

### **Sugar-Induced Chronic Inflammation: Immunometabolic Crosstalk Activation of Inflammatory Pathways**

The transition from metabolic homeostasis to chronic inflammation begins with the detection of nutrient-derived danger signals. In both immune and non-immune cells, hyperglycemia and high fructose levels are the main triggers that start several inflammatory signaling pathways that have been preserved throughout evolution

#### **The Signaling Axis of NF-κB and IKKβ**

An major regulator of sugar-associated inflammatory responses, the NF-κB signaling cycle is a well-known core route that controls the expression of numerous genes that underlie immune activation and inflammation. Under healthy conditions, NF-κB is located in the cytoplasm attached to IκB family inhibitory proteins when there is no glucose. But when nutrients are abundant and blood glucose remains high, the IκB kinase (IKK) complex is activated by both of the catalytic subunits: IKKα and IKKβ, along with the active regulating component NEMO (Kaszycki and Kim, 2025).

After activation, IKKβ will phosphorylate IκB proteins, which will then be targeted for ubiquitin-dependent degradation by the proteasomal system. When these inhibitory proteins are removed, the NF-κB p50/p65 complex can move into the cell's nucleus and come into contact with regulatory DNA sequences that cause the production of many inflammatory components, such as interleukin-6 (IL-6), interleukin-1 beta (IL-1β), and tumor necrosis factor alpha (TNF-α) (Sousa et al., 2023).

Metabolic overload also has been linked with endoplasmic reticulum (ER) stress, that additionally enhances the activity of IKK $\beta$ /NF- $\kappa$ B signaling. It is found that this pathway is chronically activated in metabolic organs and areas in the central nervous system like the hypothalamus, which disrupts insulin and leptin action networks. These changes lead to decreased energy regulation, metabolic imbalance and the production of insulin resistance (Schmidt et al., 2001).

### The NLRP3 Inflammasome and Two-Step Activation

A multiprotein intracellular complex called the NLRP3 inflammasome is responsible for identifying danger and metabolic stress signals in cells. (López-Otín et al. 2013; Sousa et al. 2023).. The 2-signal model of NLRP3 inflammasome activation: Pro-inflammatory cytokines like IL-1 $\beta$  and IL-18, which have been found to be essential elements of the inflammatory response brought on by carbohydrates, are regulated and released by this complex.

Signal 1 (Priming): Toll-like receptors, particularly TLR4, are activated by chronic hyperglycemia and gut-derived Lipopolysaccharides (LPS). This activates the NF- $\kappa$ B pathway, increasing the expression of NLRP3 and pro-IL-1 $\beta$ .

Signal 2 (Activation): Consuming a lot of sugar encourages the build-up of metabolic danger signals, including modified low-density lipoproteins (LDL), cholesterol crystals, and mitochondrial reactive oxygen species (ROS). The NLRP3 complex (NLRP3, ASC, and caspase-1) is formed as a result of these stimuli. This facilitates the breakdown of pro-IL-1 $\beta$  into its active and secretory form (Kaszycki and Kim, 2025; Sousa *et al.*, 2023).

Macrophage sensitivity of NLRP3 pathway has been demonstrated to be increased by high levels of glucose. High glucose levels cause Bone-Marrow Derived Macrophages (BMDM) to remodel their inflammatory activity, which increases IL-1 $\beta$  release in response to stimuli like nigericin or LPS (Sousa *et al.*, 2023). This molecular sensitization explains the hyper-reactive inflammatory state observed in individuals with chronic high sugar intake.

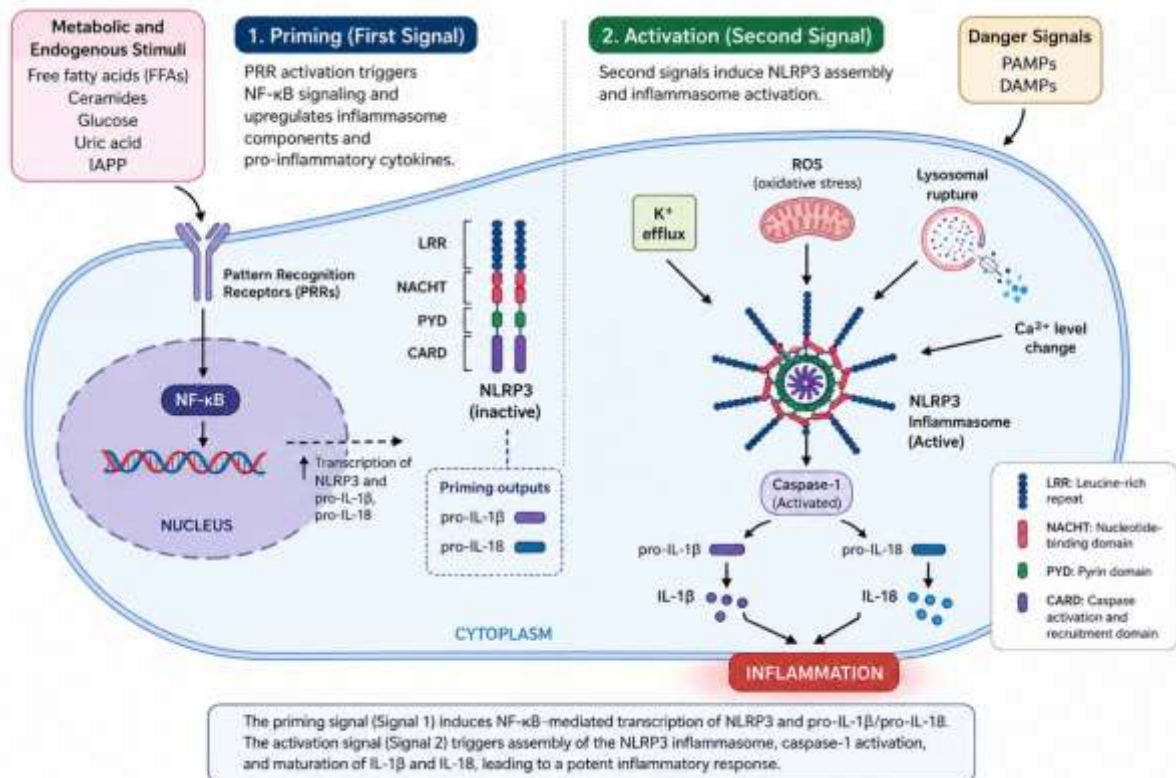


Figure 2. Metabolic and endogenous triggers activate the NLRP3 inflammasome in sugar-induced inflammation. These include free fatty acids, ceramides, glucose,

*uric acid, and islet amyloid polypeptide (IAPP) which stimulate pattern recognition receptors (PRRs). This activates NF-κB-dependent priming, which raises the level of expression of NLRP3 and pro-IL-1β and pro-IL-18. These later danger signals such as potassium efflux, production of mitochondrial reactive oxygen species (ROS), damage to lysosomes, disruptions of calcium balance, pathogen-associated and damage-associated molecular patterns (PAMPs and DAMPs) help to assemble the NLRP3 inflammasome complex and activate caspase-1. Upon activation, caspase-1 converts the inactive forms of IL-1β and IL-18 to mature active forms (IL-1β, IL-18) that will also amplify inflammation.*

Second, cellular stress responses like potassium efflux, mitochondrial ROS production, lysosomal rupture, cathepsin B release and calcium imbalance are induced by danger signals like PAMPs and DAMPs. Pro-caspase-1 is changed into active caspase-1 by these events, which also activate the NLRP3 complex. Activated caspase-1 further processes IL-1β and IL-18 into their inflammatory forms and then they are secreted from the cell, leading to inflammation (Al Mamun et al., 2021).

### **MAPK Signalling Cascade**

This is also true of the MAPK family signaling proteins, which also get activated by a high sugar diet, in addition to ERK, JNK and p38. Metabolic balance in the cell is lost, and the sugar derivative products are formed and bind to the Receptor for Advanced Glycation End-products (RAGE), which activates the kinases.

(Moreno-Fernandez et al., 2023). Additionally, the JNK signaling pathway has been shown to promote insulin resistance by increasing the serine phosphorylation of Insulin Receptor Substrate-1 (IRS-1), thereby decreasing insulin-mediated signaling. Also, regular intake of foods with a high glycemic index increases the level of insulin in circulation, which can trigger the insulin-like growth factor-1 (IGF-1) signaling pathway. IGF-1 protects cells against apoptosis, stimulates cell growth, and enhances malignant cell transformation in cells such as those from the bladder epithelium via AKT and MAPK pathway. This establishes a clear connection between the oncogenesis process and inflammation brought on by sugar (Moreno-Fernandez *et al.*, 2023).

### **Autophagy Dysfunction and Maintenance of Inflammation.**

One of the most important but often overlooked aspects of inflammation caused by sugar is the disruption of autophagy. Autophagy is a self-maintenance mechanism to recycle damaged organelles, storing pro-inflammatory proteins, including active IL-1β, to ensure the tissue damage will not increase (Sousa *et al.*, 2023). The increased autophagic proteins such as the LC3b and Beclin-1 are produced by cells under normal stress. This autophagic pathway is essentially impaired in diabetic or chronically hyperglycemic diseases. The level of LC3b and Beclin-1 in the macrophages of diabetic mice reduces dramatically, which means that the cellular cleaning system is not functioning properly (Sousa *et al.*, 2023). Because the cell is unable to clear such inflammatory mediators, this deficiency leads to the accumulation of damaged mitochondria and the chronic release of IL-1b (Sousa *et al.*, 2023). One major reason is the current meta-inflammation of the system is that inflammation does not disappear at the cellular level. These inflammatory mechanisms and their consequences are mentioned in Table 4.

**Table 4. Important Inflammatory Signaling Pathways Activated by Chronic High Sugar Intake and Their Clinical Consequences**

<b>Pathway Component</b>	<b>Triggering Mechanism</b>	<b>Principal Biological Effect</b>	<b>Health Outcome</b>	<b>References</b>

NF-κB (Nuclear Factor-kappa B)	NF-κB signaling is made possible by IKKβ activation, which encourages the breakdown of IκB inhibitory proteins.	Pro-inflammatory mediators such as TNF-α, IL-6, and IL-1β have been found to be produced at higher levels.	Low-grade systemic inflammation that persists	Baker et al. (2011); Sousa et al. (2023)
NLRP3 Inflammasome (Pyrin Domain-Containing Protein 3, Leucine-Rich Repeat, and Nucleotide-Binding Oligomerization Domain).	Sequential priming and activation by metabolic danger signals	Increased IL-1β maturation and release	Tissue damage and impaired insulin sensitivity	Sousa et al. (2023)
MAPK Family (ERK, JNK, and Related Kinases)	Activation following RAGE-mediated cellular stress responses	Increased IRS-1 serine phosphorylation that compromises insulin signaling pathways	Production of insulin resistance	Tratnjek et al. (2026)
Autophagy Pathway (LC3B-Associated Cellular Clearance Mechanism)	Suppression of autophagic activity under chronic hyperglycemic conditions	Reduced clearance of inflammatory mediators and accumulation of IL-1β	Long-term maintenance of inflammatory responses	Sousa et al. (2023)

### **Oxidative Stress–Inflammation Axis**

Sugar-induced inflammation is inextricably linked to oxidative stress. High sugar and fructose intake increases the production of free radicals, which serve as key messengers to further activate inflammation pathways, leading to a vicious cycle of cell damage.

### **Mitochondrial Dysfunction and Superoxide Overproduction**

During hyperglycemia, a major source of oxidative stress is the mitochondrial electron transport chain. When cells are exposed to high glucose, endothelial cells of large and small blood arteries make too many of the superoxide radicals (Giacco and Brownlee, 2010). This overproduction of superoxide is a triggering event at the beginning of a number of deleterious biochemical pathways. Hyperglycemia induces an increased flux in the polyol pathway, and up to 30% of the available glucose can be metabolized through this pathway in tissues such as the retina, which results in a decrease in NADPH and glutathione levels (Moreno-Fernandez *et al.*, 2023). Concurrently, superoxide promotes the synthesis of diacylglycerol (DAG), which in

turn activates several Protein Kinase C isoforms (Moreno-Fernandez *et al.*, 2023). Advanced glycation end products (AGEs) are produced more frequently as a result of oxidative stress, which also speeds up the non-enzymatic alteration of proteins.

### **Mitochondrial Dysfunction and Superoxide Generation**

Mitochondria are a significant intracellular source of oxidative stress when glucose levels are high. As glucose availability rises, more electrons flow by the electron transport chain leading to too many electrons being leaked and more development of superoxide radicals in vascular endothelial cells (Giacco and Brownlee, 2010). The increase in ROS accelerates a chain of molecular events that result in cell damage and metabolic dysfunction.

Another metabolic pathway that can be activated by excess glucose is the polyol pathway. In tissues that are highly susceptible, like the retina, a significant amount of glucose inside the cell can be consumed in this pathway, which can cause the NADPH content to drop, thereby decreasing antioxidant levels because glutathione is also depleted (Moreno-Fernandez *et al.*, 2023). At the same time, oxidative stress results to the increase of diacylglycerol (DAG) that will then cause acceleration of several Protein Kinase C (PKC) isoforms. Furthermore, ROS increases the rate of the glycation process of macromolecules in the cell, thereby enhancing the generation and accumulation of advanced glycation end products (AGEs) (Moreno-Fernandez *et al.*, 2023).

### **Signaling pathways mediated by the receptor for advanced glycation end products (RAGE) and advanced glycation end products (AGEs).**

AGEs are a sign of long-term metabolic problems brought on by excessive exposure to sugar. These substances are produced by non-enzymatic interactions between biological macromolecules including proteins, lipids, and nucleic acids and reducing sugars like glucose, fructose, and galactose (Moreno-Fernández *et al.*, 2023). Once produced, AGEs can interact with the Receptor for Advanced Glycation End Products (RAGE) and function as endogenous danger signals. Reactive oxygen species (ROS) and reactive nitrogen species (RNS) are produced when AGEs bind to RAGE, causing a strong inflammatory response that increases NADPH oxidase (NOX). This oxidative condition increases the production of RAGE by activating NF- $\kappa$ B signaling, which intensifies the inflammatory response (Moreno-Fernandez *et al.*, 2023).

Neutrophils release inflammatory enzymes like myeloperoxidase (MPO) and elastase when AGE-RAGE is activated. By promoting CD4+ T cell development toward pro-inflammatory Th1 and Th17 phenotypes, these signaling molecules can affect adaptive immunological responses and contribute to the persistence of chronic inflammatory diseases (Moreno-Fernandez *et al.*, 2023).

### **The Nrf2/ARE Defense System and its Failure**

One important cellular defense mechanism against oxidative stress is the Nrf2/ARE (antioxidant response element) pathway. As a transcription factor, Nrf2 enhances cellular defense by controlling the expression of antioxidant enzymes including catalase and superoxide dismutase (SOD) (Moreno-Fernandez *et al.*, 2023). Acute oxidative stress activates this protective pathway, but eating excessive amount of sugar over a long period of time can wear it out or damage it. Certain micronutrients, including curcumin, vitamins A and E, and polyphenols, function as ROS scavengers and can activate the Nrf2 pathway, presenting a potential approach for reducing oxidative damage caused by sugar. The interplay between Nrf2 and sirtuin 1 is essential for safeguarding mitochondrial integrity and renal function during chronic hyperglycemia (Moreno-Fernandez *et al.*, 2023). Key oxidative stress pathways and markers are summarized in Table 5.

**Table 5. Key Oxidative Stress Markers Induced by Hyperglycemia and Their Long-Term Pathophysiological Effects**

Oxidative Marker	Source/Inducer	Immediate Effect	Long-term Impact	References
Superoxide	Mitochondrial ETC (Electron Transport Chain) flux	Nuclear Factor kappa B (NF-κB) and Protein Kinase C (PKC) signaling pathways are activated.	Vascular damage	Brownlee (2001); Moreno-Fernández <i>et al.</i> (2023)
Advanced Glycation End Products (AGEs) are end products of glycation processes.	Interaction with the Advanced Glycation End Product Receptor (RAGE).	Reactive oxygen species (ROS) production.	Chronic inflammation	Tratnjek <i>et al.</i> (2026)
NOX Oxidase Enzymes	Protein Kinase C (PKC) activation	Reactive Nitrogen Species (ROS/RNS) explosion	DNA damage	Moreno-Fernández <i>et al.</i> (2023)
DAG (Diacylglycerol)	Glycolytic intermediates	PKC activation	Insulin resistance	Moreno-Fernández <i>et al.</i> (2023)

### **Adipose Tissue, Hepatic Immune Remodeling.**

Structural and cellular remodeling of metabolic organs, especially adipose tissue and liver, is one of the most significant effects of chronic sugar eating. These tissues become depositories of energy in areas of active immune interactions.

### **Adipose Tissue: Homeostasis to Meta-inflammation.**

Adipose tissue is also a fat-storage organ, and serves as a vital link between immune control and metabolic functions. It consists of a complex network of immune cells (macrophages, T cells, B cells, eosinophils and others) that collectively maintain physiological homeostasis (Wang and Wu, 2018). The adipose tissue generates a balancing effect to establish an immunological microenvironment comparable to Type 2 in a healthy organism. M2 macrophages also help in tissue regeneration via release of anti-inflammatory cytokines, such as IL-10. Visceral adipose tissue (VAT) has a large number of regulatory Tregs, which help to maintain insulin sensitivity.

### **Adipose Tissue and Hepatic Immune Remodeling**

One of the most important side-effects of chronic sugar eating is a structural and cellular remodeling of metabolic organs, particularly adipose tissue and liver. During periods of active immune interactions these tissues can become energy-storage areas.

### **Adipose Tissue: Homeostasis to Meta-inflammation**

Adipose tissue serves as a vital link between immune system activity and metabolic regulation in addition to being a location for storing energy. Its dynamic network consists of immune cells such as T cells, B cells, eosinophils and macrophages, which

mediate general physiological balance and health (Wang and Wu, 2018). In a healthy condition, adipose tissue produces elements that create a type 2 immune milieu which ensure a balanced immune response. In this situation, M2 macrophages aid in tissue healing processes by releasing anti-inflammatory cytokines like IL-10. In addition, there are many regulatory T cells (Tregs) in visceral adipose tissue (VAT) that play a crucial role in the maintenance of insulin sensitivity and metabolic homeostasis.

Sugar consumption for an extended period of time and excessive calorie expenditure greatly affect the immunological profile of adipose tissue. It is characterized by a deregulated, relative increase of pro-inflammatory lymphocytes like CD8+ T lymphocytes, CD4+ T lymphocytes (Th1 and Th17) and a relative decrease of CD4+ T lymphocytes (Tregs), which usually act as a regulating cell. Macrophages can be polarized to a pro-inflammatory state (M1) by the increased production of Monocyte Chemoattractant Protein-1 (MCP-1). In addition to forming crown-like structures around dead or dying adipocytes, these activated M1 macrophages release large amounts of pro-inflammatory cytokines like TNF- $\alpha$  and IL-6, which lead to decreased insulin sensitivity and the emergence of insulin resistance (Wang and Wu, 2018).

### **Hepatic Immune Remodeling and Fructose Metabolism**

Since the liver is the primary organ in charge of fructose metabolism, it is especially susceptible to remodeling brought on by sugar. In the liver, fructose metabolism skips the steps that slow down glycolysis. This speeds up lipogenesis and the production of lipid species that cause inflammation (Moreno-Fernández *et al.*, 2023). Kupffer cells (KCs) are specialized macrophages that reside within the liver. They are located in the sinusoids and clear products from the gut without causing an immune response (Vasudevan *et al.*, 2024). But eating a lot of fructose changes the biology of KCs:

**Pentose Phosphate Pathway (PPP) Regulation:** Fructose regulates the PPP in Kupffer cells, inducing a phenotype that is simultaneously inflammatory and resolution-oriented. **Monocyte Recruitment:** Chronic fructose intake results to lessen in the proportion of yolk-sac-derived resident KCs and an increase in transitioning monocytes and infiltrating monocytes from the bone marrow. **Cytokine Cascade:** The condition can proceed from simple fat accumulation in the liver (steatosis) to non-alcoholic steatohepatitis (NASH) and finally fibrosis. Recruited macrophages release IL-1 $\beta$ , TNF- $\alpha$ , and IL-6, which activate hepatic stellate cells (Moreno-Fernández *et al.*, 2023; Vasudevan *et al.*, 2024).

### **The Liver-Adipose Axis in Metabolic Stress**

During sugar-induced stress, there is a strong crosstalk between adipose tissue and liver during sugar-induced stress. Dysfunctional adipose tissue releases pro-inflammatory adipokines and fatty acids into the portal circulation that directly activates the hepatic Kupffer cells (Wang and Wu, 2018; González *et al.*, 2023). On the other hand, the liver produces inflammatory mediators like IL-11 and various hepatocytes, which can aggravate the inflammation of adipose tissue (González *et al.*, 2023). This inter-organ axis ensures that inflammatory signals are disseminated throughout the body rather than remaining localized. This inter-organ axis makes sure that inflammatory signals are spread throughout the body instead of staying in one place. The immune remodeling of metabolic tissues is summarized in Table 6.

**Table 6. Immune Cell Remodeling in Adipose and Hepatic Tissues During Sugar-Induced Metabolic Dysfunction**

Cell Type	Healthy State	Sugar-Induced State	Key Signaling Molecules	References
T Cells	By promoting anti-	Increased	Interleukin-	Trantjek <i>et</i>

	inflammatory reactions, regulatory T cells (Tregs) and T helper 2 (Th2) cells are essential for maintaining immunological balance.	infiltration of T helper 1 (Th1), T helper 17 (Th17), and CD8 <sup>+</sup> cytotoxic T cells that cause inflammation	17 (IL-17) and interferon-gamma (IFN- $\gamma$ )	<i>al.</i> (2026)
<b>Kupffer Cells</b>	Immunotolerant liver-resident macrophages involved in clearance of microbial products without excessive immune activation	Activated Kupffer cells contributing to hepatic inflammation, fibrosis, and progression of metabolic dysfunction	C-C Motif Chemokine Ligand 2 (CCL2) and Interleukin-1 beta (IL-1 $\beta$ )	Vasudevan <i>et al.</i> (2024)
<b>Innate Lymphoid Cells</b>	Tissue homeostasis and anti-inflammatory immune modulation are supported by group 2 innate lymphoid cells (ILC2).	Pro-inflammatory innate lymphoid cell subsets promoting metabolic inflammation	Interleukin-33 (IL-33)	Wang and Wu (2018)

### **Gut Microbiome and Metabolic Endotoxemia.**

The first point of interaction of dietary sugar is the gastrointestinal tract. Metabolic endotoxemia is a disorder brought on by continuous acute high-sugar diets (HSD), which upset the gut microbiota's equilibrium and jeopardize the integrity of the intestinal barrier.

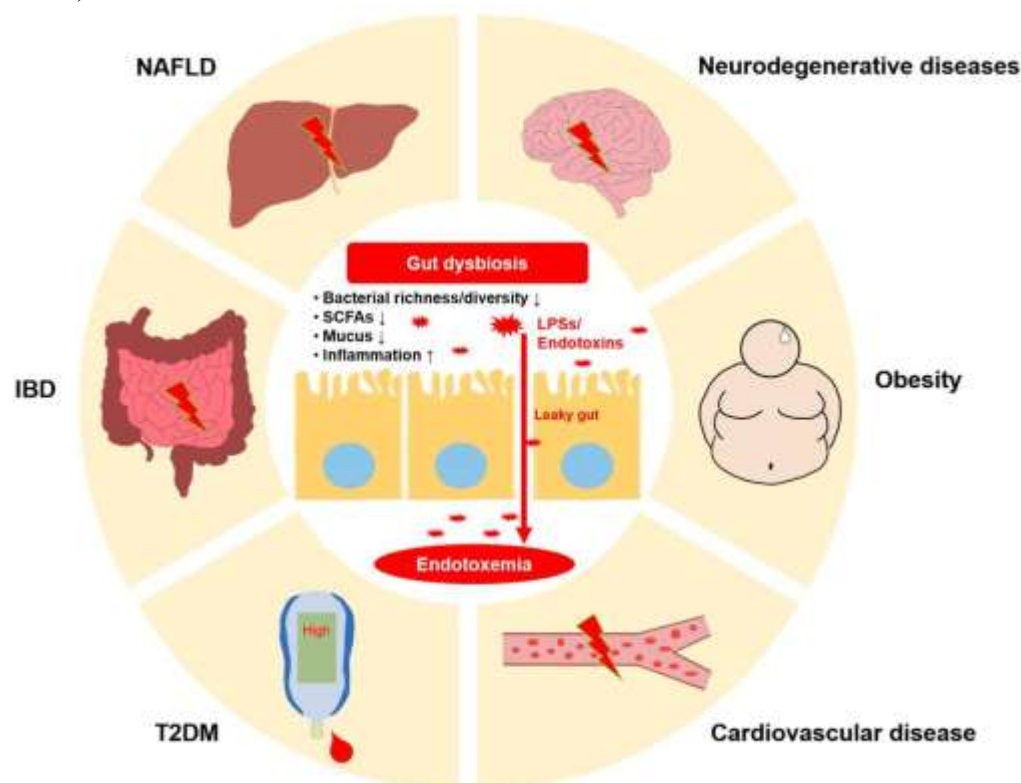
### **Barrier Failure and Mucus Degradation**

The mucus layer which consists mainly of the protein Muc2, lines the intestinal epithelium and prevents direct contact between gut microbes and epithelial cells. Consumption of large quantities of simple sugars like sucrose and fructose has been demonstrated to significantly reduce the expression and decrease the thickness of the mucus layer (Fajstova *et al.*, 2020). This barrier dysfunction allows potentially harmful microbes to approach the intestinal wall more closely. High-sugar diets (HSDs) are linked to gut microbial imbalance, which is characterized by an increase in bacteria like *Candida* and *Escherichia coli*, a decrease in beneficial bacteria that produce short-chain fatty acids (SCFA), and damage to the integrity of the intestinal tight connection (Fajstova *et al.*, 2020; Moreno-Fernández *et al.*, 2023). Because tight junction proteins are broken down in this state, bacterial elements are translocated into the circulation.

### **LPS Translocation and TLR4 Activation**

Lipopolysaccharide (LPS), a structural element of Gram-negative bacteria's outer membrane, can translocate into the portal circulation when sugar-induced disruption of gut barrier function occurs. A two- to three-fold increase in circulating LPS levels is usually indicative of metabolic endotoxemia, which is caused by this mechanism (Fajstova *et al.*, 2020). After entering the bloodstream, lipopolysaccharide (LPS) attaches itself to the immune cells' TLR4/CD14 complex, starting two main signaling

pathways. Pro-inflammatory cytokines like TNF- $\alpha$  and IL-6 are produced quickly as a result of the MyD88-dependent pathway's activation of NF- $\kappa$ B and mitogen-activated protein kinase (MAPK) signaling. On the other hand, type I interferon expression is induced by the MyD88-independent route via IRF3 activation. Systemic inflammation is greatly exacerbated by persistent low-level TLR4 stimulation, which has also been linked to the exacerbation of diseases like colitis and metabolic syndrome. Additionally, research using TLR4-deficient mice shows resistance to the inflammatory effects of high-sugar diets, highlighting the crucial role this receptor plays in mediating these reactions (Fajstova *et al.*, 2020; Moreno-Fernández *et al.*, 2023).



**Figure 3.** Gut dysbiosis and metabolic endotoxemia linked to excessive sugar consumption involve significant alterations in intestinal microbial balance. Consuming large amounts of sugar can disrupt the gut microbiota, reduce the number of bacteria that produce short-chain fatty acids (SCFA), damage the integrity of the intestinal epithelium, and compromise the mucus barrier. As a result, gut permeability increases, permitting lipopolysaccharides (LPS) and other bacterial endotoxins to translocate into the bloodstream, thereby contributing to metabolic endotoxemia. Numerous chronic problems, such as non-alcoholic fatty liver disease (NAFLD), inflammatory bowel disease (IBD), type 2 diabetes mellitus (T2DM), cardiovascular disorders, obesity, and neurodegenerative diseases, have been associated with this process, which encourages broad systemic inflammation.

### Short-Chain Fatty Acids' (SCFAs') Function

Microbial fermentation of dietary fiber produces short-chain fatty acids (SCFAs), primarily acetate, propionate, and butyrate. They are very important for keeping the intestinal barrier strong because they give colonocytes energy and help Regulatory T cells (Tregs) in the gut become different (Fajstova *et al.*, 2020). High sugar intake takes away this important anti-inflammatory "brake" by lowering the number of bacteria that make SCFAs. When SCFAs are lost, the gut becomes more permeable, and the immune system becomes more pro-inflammatory. On the other hand, eating a lot of fiber or taking probiotics can help bring SCFA levels back to normal and reduce inflammation caused by sugar.

## Metabolomics and Systemic Shifts

Targeted metabolomics studies in humans have demonstrated that high-fructose diets induce significant alterations in plasma metabolites. In particular, mitochondrial  $\beta$ -oxidation is inhibited, and lipid peroxidation is heightened, as indicated by alterations in acyl carnitine and lysophosphatidylcholine concentrations (Moreno-Fernández *et al.*, 2018). These metabolic changes are stronger when fructose is given in high-fructose syrup instead of whole fruit. This suggests that the type of food and how quickly it is given affect how strong the inflammatory response is (Moreno-Fernández *et al.*, 2018). The impact of high sugar intake on gut microbiota and inflammation is summarized in Table 7.

**Table 7. Effects of High Dietary Sugar Consumption on Gut Microbiota Balance and Inflammatory Responses**

Microbiota Component	Effect of High Sugar Intake	Inflammatory Consequence	Reference
<b>Muc2 / Mucus Layer</b>	Reduced mucus production and impaired barrier integrity	Enhanced bacterial translocation and mucosal inflammation	Fajstova <i>et al.</i> (2020)
<b>LPS (Lipopolysaccharide)</b>	Increased circulating endotoxin levels	Activation of the pro-inflammatory reactions linked to the TLR4/NF- $\kappa$ B signaling cascade.	Tratnjek <i>et al.</i> (2026)
<b>SCFAs (Short-Chain Fatty Acids)</b>	Depletion because of altered microbial metabolism	Loss of intestinal barrier function and increased inflammation	Fajstova <i>et al.</i> (2020)
<b>Escherichia coli (E. coli)</b>	Increased abundance under dysbiotic conditions	Major source of LPS contributing to systemic inflammation	Fajstova <i>et al.</i> (2020)

## Molecular Interventions between Dietary Sugar and Premature Aging

Dietary sugar overconsumption has become more evidently considered a severe environmental cause of biological aging. High glucose and fructose exposure causes molecular alterations that do overlap significantly with the known hallmark of aging, while being generally associated with metabolic disorders like obesity and type 2 diabetes. Prolonged sugar intake promotes chronic hyperglycemia, insulin resistance, mitochondrial overload and oxidative stress, and produces a biochemical environment that facilitates progressive cellular harm. These processes affect the genomic stability, proteostasis, telomere stability, and nutrient-sensing mechanisms, all of which play a major role in aging patterns (López-Otín *et al.*, 2013). Unlike acute metabolic changes, a longer duration of dietary sugar triggers molecular stress that is low-intensity but chronic over time ultimately compromising cellular resilience. Therefore, tissues exposed for a longer time to glycemic and fructose stress condition exhibit impaired tissue functioning which can be compared to an accelerated aging phenomenon observed in phenotype. Glycation reactions, accumulation of DNA damage, damage to mitochondria, and senescence of the cell are the processes at the molecular level involved in this relationship. Combined, those processes create a high demand for sugar in the diet and make it a significant risk factor in metabolism, but also a systemic cause of biological aging.

### **AGEs, or advanced glycation end products**

An important molecular connection between food and ageing is the production of advanced glycation end products (AGEs). The Maillard reaction is a non-enzymatic reaction that occurs between free amino groups of proteins, lipids or nucleic acids and reducing sugars (Verzijl *et al.*, 2000; Singh *et al.*, 2001). Chronic hyperglycemia leads to a much greater response which leads to the accumulation of irreversibly modified macromolecules (Brownlee, 2001). Fructose is more reactive than glucose because of its open chain structure and rapid rate of intracellular metabolism, which increases the rate of the formation of AGEs (Suarez *et al.*, 1989). These glycation reactions change the structure and function of proteins, inhibit enzyme activity, cross-link the long-lived proteins of the extracellular matrix (e.g., collagen, elastin), and ultimately impair tissue elasticity and functional integrity (Monnier *et al.*, 2005; Paul and Bailey, 1996). In addition to structural impacts, AGEs initiate down stream signaling cascades through binding with the Receptor for Advanced Glycation End Products (RAGE). RAGE activation can lead to sustained inflammatory responses, increased oxidative stress and activation of NF- $\kappa$ B signaling (Schmidt *et al.*, 1999). This chronic inflammatory stimulation is one of the mechanisms that contribute to increased vascular stiffness, renal dysfunction and neurodegenerative changes that commonly occur in the course of age. Notably, AGE levels build up proportionately with age but are greatly escalated in hyperglycemic states implying that excessive sugar consumption enhances inherent aging processes (Brownlee, 2001). Endogenous AGEs produced under the influence of processed, high-sugar foods are also added to the dietary AGEs, which enhance systemic burden and oxidative stress (Uribarri *et al.*, 2010). Therefore, glycation is a key biochemical intermediary between chronic exposure to sugar and structural tissue aging and inflammatory degeneration.

### **Genomic Instability and DNA Damage**

Genomic instability is widely recognized as an aging hallmark, which is marked by the presence of accumulated DNA damage and the inability to repair it (López-Otín *et al.*, 2013). Increased sugar consumption promotes genomic instability through processes brought on by oxidative stress. Sustained hyperglycemia increases mitochondrial electron transport chain activity, resulting in excessive generation of superoxide and other reactive oxygen species (ROS) (Nishikawa *et al.*, 2000). High levels of ROS damage nucleic acids and produce lesions, including 8-oxo-deoxyguanosine, as well as single and double strand breaks. Additionally, the glycoxidation reactions directly alter the DNA bases and reduce the genomic fidelity. By quickly depleting intracellular ATP and increasing the generation of uric acid, which increases mitochondrial oxidative stress and inflammatory signaling, increased fructose metabolism may further increase genomic stress (Johnson *et al.*, 2009). The chronic DNA damage triggers DNA damage response pathways that engage the p53 and the ATM/ATR signaling pathways that may induce cell cycle arrest, apoptosis, or senescence (Vousden and Prives, 2009). Although the mechanisms have protective functions in the short term, prolonged activation leads to dysfunction of tissues and stem cell exhaustion. The combination of damaged DNA repair ability along with sustained attack by oxidative agents leads to progressive genomic instability promoting aging phenotypes. In this way, the high intake of sugar leads to gradual genomic degradation that is a hallmark of tissues aged at the expense of premature aging.

### **Cellular Senescence and Telomere Shortening**

Telomeres function as defensive structures located at the last of chromosomes, preserving genomic stability throughout successive rounds of cell division. Replicative cycles are necessary for telomere shortening, which is a natural process that is accelerated by oxidative stress and persistent inflammation (Blackburn *et al.*,

2015). Higher levels of fasting glucose and insulin resistance have been related with a shorter leukocyte telomere length, indicating that there is a correlation between the dysregulation of metabolism and increased cellular aging (Zhao *et al.*, 2013). Oxidative lesions occur especially in telomeric DNA due to its large guanine content and consequently sensitive to the lesions caused by the ROS (von Zglinicki, 2002). Chronic hyperglycemia augments oxidative load, thus surpassing typical replicative shortening.

Telomeres shorten to critically short lengths and the cell become permanently growth arrested, a condition called cellular senescence. The term "senescence-associated secretory phenotype" (SASP) refers to the pro-inflammatory profile and variety of bioactive molecules secreted by senescent cells, such as cytokines, chemokines, and extracellular matrix-degrading enzymes (Coppé *et al.*, 2010; Tchkonina *et al.*, 2013). Accumulation of senescent cells interferes with tissue micro-environment, regenerative ability, and spreads chronic inflammation (Baker *et al.*, 2011). The increased sugar intake, which facilitates oxidative stress and metabolic dysfunction, increases the rates of telomere shortening and senescence accumulation, thus promoting tissue aging and predisposing age-related diseases. Hence, telomere erosion and senescence are important mechanistic intermediate outcomes between exposure to dietary sugars and systemic biological aging (Epel *et al.*, 2004).

### **Mitochondrial Aging**

The process of aging and the deterioration of the metabolism heavily rely on mitochondrial dysfunction. Persistent exposure to elevated glucose or fructose levels exerts a prolonged energetic strain on the mitochondria, specifically within the hepatocytes, adipocytes, and skeletal muscle cells (Bhatti *et al.*, 2017). High substrate availability increases electron transport chain activity and high ROS generation and reduces oxidative phosphorylation efficiency. This oxidative load is damaging over time mitochondrial DNA, proteins, and membrane lipids, leading to impairment of bioenergetic capacity and instigating fragmentation of mitochondria (Wallace, 2005). AMP-activated protein kinase (AMPK) and sirtuin signaling pathways are two important regulatory systems involved in mitochondrial maintenance that can be disrupted by high sugar consumption. AMPK repression and NAD<sup>+</sup> depletion suppress SIRT1-mediated mitochondrial biogenesis and mitochondrial stress resistance (Cantó and Auwerx, 2009; Hardie *et al.*, 2012). The process of fructose-induced ATP loss and uric acid production worsens oxidative stress and endothelial dysfunction further (Johnson *et al.*, 2013). The effects of these combined causes decrease the adaptability of mitochondria and increase metabolic inflexibility, which is an aging characteristic of tissues (López-Otín *et al.*, 2013). Sustained exposure to dietary sugar activates the aging process of the mitochondrion, reduces the efficiency of cellular energy use, and leads to the exposure to oxidative damage.

### **Overlap with the Hallmarks of Aging**

Numerous well-known signs of aging, such as genomic instability, telomere shortening, mitochondrial dysfunction, epigenetic changes, compromised proteostasis, and dysregulated nutrient-sensing pathways, are linked to excessive dietary sugar intake (López-Otín *et al.*, 2013). High-sugar conditions that activate the insulin and mTOR signaling pathways in a chronic fashion resemble excessive nutrient conditions that cause shortened lifespan and autophagy deficiencies (Fontana *et al.*, 2010; Laplante and Sabatini, 2012). During the same time, crosslinking of proteins due to glycation causes impairment of proteostasis, and oxidative stress escalates the damage of DNA and mitochondria (Mizushima and Komatsu, 2011). These related pathways create a network of reinforcement where metabolic overload increases the rate of intrinsic aging (Bonomini *et al.*, 2010). Collectively, the evidence supports the idea that dietary sugar is a strong regulator of molecular aging mechanisms. Excessive

sugar consumption can alter the biological aging pathway to early functional impairment and predisposition to diseases by favoring oxidative stress, chronic inflammation, glycation, and mitochondrial aging (Coppé *et al.*, 2010).

## Genomics, Epigenetics, and Transcriptomic Consequences

### DNA Methylation Changes

DNA methylation is a heritable epigenetic modification that can also be reversed. It affects gene expression, particularly at the cytosine residue of CG dinucleotides. Differences in the methylation status of genes involved in the response to oxidative stress, lipid metabolism, insulin signaling, and inflammation have been linked to a high sugar diet. The methylation of gene promoters caused by a high sugar diet can be affected by the acceleration of the mTOR signaling pathway. As indicated in the study on personalized nutrition, food factors, particularly sugar intake, could cause particular methylation modifications, which could affect metabolic traits (Hussain *et al.*, 2026). It has been shown that PPAR $\gamma$ , SREBP-1c, and inflammatory gene pathways have abnormal methylation patterns in metabolic disorders. Methylation has been considered to have an essential role in the pathogenesis of metabolic disorders by enhancing the rate of lipid production and the level of inflammation, while lowering metabolic flexibility. The development of short-chain fatty acids (SCFAs) has been found to be affected by sugar-induced dysbiosis in the gut microbiome, as shown in the study by Li *et al.* (2022). The altered composition of the gut microbiome may indirectly influence methylation status by altering the levels of SCFAs, which in turn regulate the activity of DNA methyl transferase (DNMT) by butyrate. The diet-microbiome-methylation axis may utilize epigenetic instability to link the development of metabolic disorders with the consumption of excessive sugar.

Also, Nicoletti *et al.* (2024) highlight the fact that non-coding RNAs and DNA methylation are closely linked, which in turn increases the severity of metabolic disorders. In fact, it has been established that pro-inflammatory genes are hypomethylated, whereas insulin-sensitive genes are hyper methylated in the phase of NAFLD and obesity. According to Park *et al.* (2017), nutritional factors can accelerate the "epigenetic clock" in the process of aging; in this regard, bad nutritional habits, i.e., the consumption of a lot of sugar in the diet, can accelerate the process of aging through accumulation of methylation changes. Representative DNA methylation changes are discussed in Table 8.

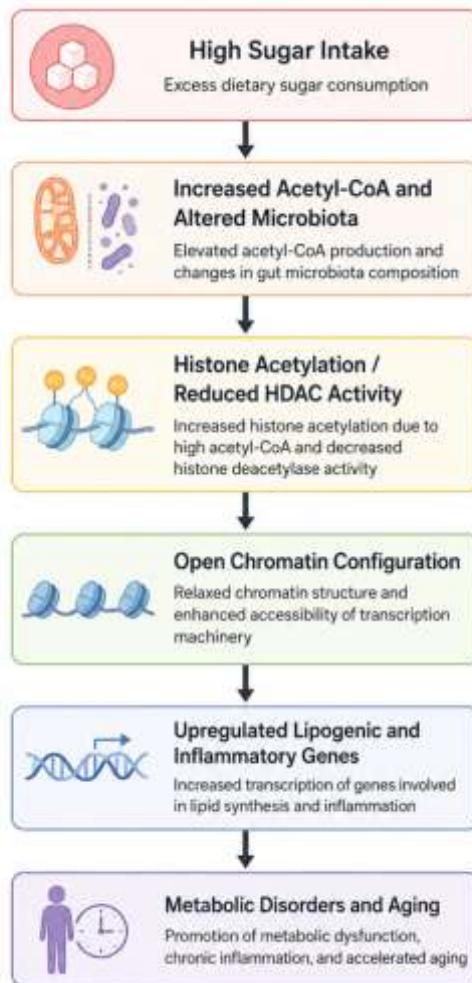
**Table 8. Representative Epigenetic DNA Methylation Alterations Induced by High Sugar Intake and Their Metabolic Consequences**

Gene/Pathway	Methylation Status	Biological Effect	Associated Condition	Reference
PPAR $\gamma$ (Peroxisome Proliferator-Activated Receptor Gamma)	Hypermethylation	decreased sensitivity to insulin and compromised development and function of adipocytes	Type 2 Diabetes (T2D)	Hussain <i>et al.</i> (2026)
Sterol Regulatory Element-Binding Protein-1c, or SREBP-1c	Hypomethylation	Increased lipogenic gene expression and hepatic lipid accumulation	NAFLD, or nonalcoholic fatty liver disease	Nicoletti <i>et al.</i> (2024)
TNF- $\alpha$ (Tumor	Hypomethylation	Elevated pro-	Metabolic	Park <i>et al.</i>

<i>Necrosis Factor-<math>\alpha</math></i> )		inflammatory cytokine output driving persistent inflammatory signaling	Syndrome	(2017)
<b>IRS1</b> ( <i>Insulin Receptor Substrate-1</i> )	Hypermethylation	Impaired insulin signaling and decreased cellular glucose uptake	Insulin Resistance	Hussain <i>et al.</i> (2026)
<b>Clock Genes</b>	Dysregulated Methylation	Disruption of circadian regulation and metabolic homeostasis	Premature Aging and Metabolic Dysfunction	Park <i>et al.</i> (2017)

### Chromatin Remodeling and Histone Alterations

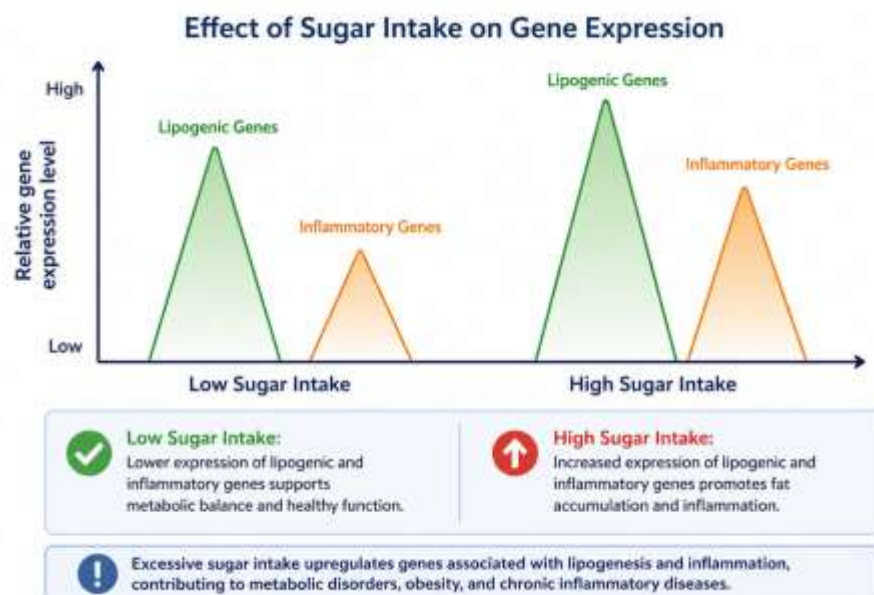
Histone alterations enable dynamic and quick control over chromatin accessibility, whereas DNA methylation offers long-term gene suppression or activation (Bannister and Kouzarides, 2011). A crucial substrate for histone acetyltransferases (HATs), intracellular acetyl-CoA levels are raised by high sucrose availability. Transcriptional activation is enhanced by increased histone acetylation at the promoters of inflammatory and lipogenic genes (Wellen *et al.*, 2009). Microbial metabolites like butyrate act as histone deacetylase (HDAC) inhibitors, according to Li *et al.* (2022). Reductions in butyrate levels weaken HDAC inhibition, which changes chromatin states in metabolic tissues when diets high in sugar disturb beneficial microbial societies. Gene expression that promotes inflammation is favored by this imbalance. Furthermore, the way that histone changes combine with non-coding RNAs to fine-tune metabolic gene networks is covered by Nicoletti *et al.* (2024). For example, transcriptional dysregulation in obesity may be exacerbated by histone acetylation at positions that regulate adipogenesis and gluconeogenesis (Zhao *et al.*, 2010). Histone remodeling caused by nutrients may take part in the development of low-grade inflammation and genomic instability, which are markers of aging, as suggested by Park *et al.* (2017). As an outcome, dietary sugar influences chromatin architecture and functions as an epigenomic modulator in addition to an energy source. These epigenetic mechanisms are illustrated in Figure 4.



**Figure 4.** *High sugar intake promotes epigenetic remodeling through increased acetyl-CoA levels and altered microbiota, resulting to enhanced histone acetylation, open chromatin structure, and upregulation of lipogenic and inflammatory genes, lastly play role to metabolic disorders and aging.*

### Transcriptomic Alterations

Dietary sugar directly alters transcriptome landscapes in addition to epigenetic markers. Particularly in NAFLD, fructose has been demonstrated to alter hepatic gene expression, fostering pathways associated with oxidative stress, de novo lipogenesis, and fibrosis (Fructose-related transcriptome modulation in NAFLD, 2021). Genes supporting mitochondrial biogenesis and insulin responsiveness are downregulated, while genes involved in fatty acid synthesis (like FASN), inflammatory mediators (like IL-6), and oxidative stress regulators are upregulated, according to transcriptomic analysis-coding RNAs, including long non-coding RNAs (lncRNAs) and microRNAs (miRNAs), are important mediators of sugar-induced metabolic imbalance, according to Nicoletti *et al.* (2024). For example, dysregulated miRNAs can promote adipocyte hypertrophy or inhibit the expression of the insulin receptor. The unique transcriptome responses to nutrition are influenced by genetic background and pre-existing epigenetic states, as further shown by Hussain *et al.* (2026). Multi-omics profiling is used in personalized nutrition approaches to predict who is more vulnerable to changes in gene expression brought on by sugar. These transcriptomic alterations are illustrated in Figure 5.



**Figure 5.** An illustration of how increased sugar consumption affects the expression of inflammatory and lipogenic genes. Low dietary sugar intake is associated with decreased activation of genes related to inflammation and lipid production, maintaining appropriate metabolic balance, as the image illustrates. On the other hand, consuming too much sugar increases the expression of genes linked to inflammation and fat formation. Chronic inflammatory diseases, metabolic dysregulation, and obesity may all be exacerbated by this elevated gene activity. Overall, the graphic highlights how excessive sugar intake negatively affects metabolic balance and cellular function.

### Genetic Susceptibility and Nutrigenomics

Nutrigenomics includes the relationship between dietary responses and genetic variation. Some genes can affect the responses of individuals to a high sugar diet. The effect of single nucleotide polymorphisms (SNPs) on genes controlling glucose homeostasis and lipid metabolism on epigenetic sensitivity to food is covered by Hussain et al. (2026). In addition to the genetic predisposition, host-microbiome interactions are also affected by genetic predisposition, as shown by Li et al. (2022). Genetic differences are ways in which microbial ecology can be changed through altered epigenetic signaling pathways. Nicoletti et al. (2024) underscore the interplay between non-coding RNAs, DNA methylation and genetic susceptibility. For instance, if they're exposed to excess sugars, they may have more methylation changes, which could speed up the disease process. According to Park et al., (2017), the alteration of epigenetic mechanisms by the diet can accelerate or decelerate ageing depending on the genetic background.

### Integrative Systems Biology Perspective

Metabolic disorders, their diet induced pathophysiology, and the aging process are not captured by a single biomolecular layer, the genome or the metabolome, for example (Hasin et al., 2017). One of the key components of systems biology is multi-omics integration, a technique that involves the study of multiple layers of biology (e.g. microbiome, metabolome, proteome and phenome) simultaneously (Bzdok *et al.*, 2018). In addition to demonstrating the mechanisms of disease leads to from disrupted cellular networks due to dietary factors such as high sugar, this approach links environmental exposures to host effects, including immune responses and aging (Sonnenburg and Bäckhed, 2016; Wishart, 2016).

## **Integration of Multi-Omics**

The integration of various high-throughput molecular data sources to understand complicated life functions and disease mechanisms is called to as multi-omics integration (e.g., genomics, metagenomics, metabolomics, and proteomics). The integrated approaches enable a better understanding of the physiology and pathology as a whole. This is achieved by uncovering the associated molecular patterns and causal relationships across various levels, as opposed to focusing on individual omics patterns.

For example: Metagenomics sequencing and targeted metabolomics are combined in a mouse model of metabolic dysfunction-associated steatotic liver disease (MASLD) to show how a Mediterranean diet affects gut microbiota and liver metabolism. This study showed that by modifying the composition of the gut microbiota and metabolic outputs, a Mediterranean diet high in complex carbohydrates, polyphenols, and unsaturated fatty acids can lower inflammation and hepatic steatosis.

Similarly, the Mediterranean diet intervention was particularly enriched in beneficial taxa (*Prevotella* and *Muribaculum*) relative to both high-fat diet control and normal chow-switched mice, as accompanied by enhanced levels of protective short-chain fatty acids (SCFAs) and reduced pro-inflammatory indicators (ALT, AST, IL-6 and TNF- $\alpha$ ) compared with the advanced fibrosis groups (Wang *et al.*, 2025). As a representative multi-omics crosstalk of host and gut microbiome axis (i.e., gut-liver axis), this coordinated adaptation implies that systemic health benefits of diet may rely not only on host metabolic control but also on the adaptive impact of gut microbial metabolite signaling.

Protein quantification, or targeted/untargeted metabolomics for metabolite profiling, high-throughput sequencing techniques such as 16S or shotgun metagenomics, and statistical integration techniques such as correlation networks, partial least squares regression, and latent factors are often used in this approach (Integrative Human Microbiome Project Consortium, 2019). Researchers can map dietary impacts to functional pathways using these integrated layers. For example, they can connect inflammatory networks, microbial dysbiosis, and dysregulated lipid metabolism, which all lead to metabolic diseases (Turnbaugh *et al.*, 2006).

We conducted multi-omics analyses within the Integrative Multi-omics Study of Sleep Apnea, including gut microbiome profiling, metabolomics, proteomics, and Phenome-wide Association Study, using a bidirectional Mendelian randomization approach to examine causal relationships between gut microbial taxa, inflammation-related proteins, and sleep apnea phenotypes (Davey Smith and Hemani, 2014). Although not diet-specific, this work shows how phenotypic outcomes, circulating proteome, genetics, and microbial composition can be combined to identify biologically relevant targets (such as the TIMP4 protein) and interactions between microbiome mediators. Among the main advantages of multi-omics integration are:

**Mechanistic pathway identification:** Combined omics bring to light immunological and metabolic changes (such as lipid and energy metabolism, SCFA production) that single layers would overlook (Sonnenburg and Bäckhed, 2016). **Biomarker discovery:** Integrated signatures from various omics layers, including taxa related to inflammatory profiles or metabolomics changes associated with early metabolic failure, improve disease prediction and classification (Wishart, 2016; Menni *et al.*, 2013).

Multi-omics in systems modeling enables the construction of causal molecular networks. These include associations between food, microbial metabolites, host metabolic gene expression, and inflammatory markers (Koh *et al.*, 2016).

## **Convergence of Metabolic and Aging Pathways**

Multi-omics studies have shown that aging, metabolic disease, and chronic inflammation have common molecular signatures. As we age, our genes, metabolism,

immune system, and microbiome change simultaneously. This leads to “inflammaging,” A persistent, mild inflammatory condition linked to impaired metabolism and the development of various diseases (Zhang *et al.*, 2024). Multi-omics studies integrating microbiome sequencing and metabolomics have shown that aged mice have increased pathobionts, increased endotoxin levels, and differential metabolites of secondary bile acids, linking microbial dysbiosis to systemic inflammation (Wang *et al.*, 2025).

Dietary factors also have a role in mediating this. A diet high in sugar and saturated fats may produce dysbiosis, change the gut flora, and impact bile acid and lipid homeostasis. This may lead to an increase in pro-inflammatory metabolites and a decrease in anti-inflammatory SCFAs, further exacerbating insulin resistance and metabolic dysfunction (Li *et al.*, 2025). In the non-gut context, "aging archetypes" with specific metabolic and inflammatory profiles are defined by multi-omics analysis, such as transcriptomics, lipidomics, and phenotyping. This highlights the connection between immune system modulation, energy metabolism, and aging-related diseases (Chen *et al.*, 2025).

In conclusion, multi-omics studies demonstrate that the biological networks influenced by nutritional, microbial, metabolic, and inflammatory processes are shared between metabolic diseases and aging. This systems biology perspective highlights the importance of lifestyle decisions, such as sugar consumption, in regulating aging and metabolism (Wang *et al.*, 2025). These integrative multi-omics relationships are summarized in Table 9.

**Table 9: Multi-Omics Insights Linking Diet, Microbiome, and Aging**

Omics Layer	Biological Signal	Outcome	Example Finding	References
Metagenomics	Alterations in gut microbiome composition and Diversity	Influence on metabolic health and inflammatory regulation	Enrichment of <i>Prevotella</i> species associated with dietary patterns and metabolic responses	Wang <i>et al.</i> (2025)
Metabolomics	Variations in short-chain fatty acids (SCFAs) and metabolic intermediates	Reduced systemic inflammation and improved metabolic homeostasis	Increased SCFA production linked with improved gut and metabolic function	Wang <i>et al.</i> (2025)
Proteomics	Expression of inflammatory and metabolic regulatory proteins	Identification of biomarkers associated with chronic disease risk	Tissue Inhibitor of Metalloproteinase-4 (TIMP4) associated with sleep apnea and metabolic dysfunction	Li <i>et al.</i> (2025)
Phenome	Changes in body composition and physiological characteristic	Improved disease prediction and metabolic risk assessment	Bioelectrical impedance markers linked with obesity and metabolic disease prediction	Li <i>et al.</i> (2025)

### Implications for Therapy and Prevention

A growing body of data from mechanistic and epidemiological studies indicates that consuming too much sugar is not just a lifestyle-related risk factor but also directly contributes to the development of metabolic dysfunction, chronic inflammation, and

accelerated aging processes (Srouf *et al.*, 2019). According to molecular research, consuming too many free sugars—especially fructose and sucrose—activates biological pathways associated with oxidative stress, insulin resistance, mitochondrial damage, and the build-up of advanced glycation end products (AGEs). These interrelated processes are linked to cardiovascular disease, cancer, and neurological illnesses in addition to contributing to the development of metabolic syndrome (Hotamisligil, 2017; Fontana *et al.*, 2010).

In order to translate this molecular insight into a practical approach for clinical and public health medicine, pharmaceutical, nutritional, and personalized therapies must be combined. Following a review of meta-analyses, the comprehensive umbrella review published in the BMJ in 2023 concluded that a high dietary intake of free sugar is significantly associated with an increased risk of obesity, type 2 diabetes, cardiovascular disease, and all-cause mortality (Huang *et al.*, 2023). Most importantly, these associations were strongest for added sugar-containing beverages, which rapidly increase postprandial insulin and glucose concentrations, accelerating oxidative stress and glycemic fluctuations (Y. Huang *et al.*, 2023). Via cellular senescence and epigenetics, these systemic derangements create an internal environment that is pro-inflammatory and accelerates biological aging.

### **Dietary Interventions**

The most important and scalable approach to mitigate sugar-induced molecular damage is through dietary modifications. As recommended by global health organizations, restricting free sugar to below 10% of total daily calorie intake reduces the demand for insulin and mitigates postprandial hyperglycemia. Biochemically, restriction of sugar intake will suppress the development of reactive oxygen species in mitochondria, block the acceleration of the polyol pathway, inhibit the development of AGEs, and reduce de novo lipogenesis in the liver (Giacco and Brownlee, 2010; Brownlee, 2001; Softic *et al.*, 2020; Stanhope, 2016).

### **The Molecular Effects of Excessive Sugar Consumption**

Large quantities of glucose and fructose can disrupt the normal metabolic regulation via glycolytic and lipogenic cycle resulting to oxidative stress, lipid accumulation, insulin resistance and chronic inflammation. Consuming sugar can also result in modified glycation end products (AGEs), which activate inflammatory signaling pathways including NF-κB and speed up the receptor for better glycation end products (RAGE). These molecular level derangements are implicated in endothelial dysfunction, inflammation of adipose tissue, metabolic instability and chronic metabolic diseases (Hotamisligil, 2017; Brownlee, 2001).

### **Evidence-Based Dietary Patterns**

The protective mechanisms that show the effectiveness of dietary approaches in stabilizing glycemic indices and decreasing inflammation.

**Table 10. Evidence-based dietary interventions targeting sugar-induced metabolic dysfunction**

<b>Dietary Pattern</b>	<b>Mechanistic Target</b>	<b>Biological Outcome</b>	<b>References</b>
<b>Reduced Free Sugar Intake</b>	Decreased generation of advanced glycation end products (AGEs) and increased insulin sensitivity.	Improved glycemic control and reduced metabolic stress	BMJ (2023)
<b>Mediterranean</b>	Nuclear Factor-kappa B	Reduced	Cohort studies

<b>Diet</b>	(NF-κB) signaling is suppressed when antioxidant enzyme activity is elevated.	cardiovascular disease risk and improved longevity	and randomized controlled trial (RCT) evidence
<b>High-Fiber Diet</b>	Enhanced glucagon-like peptide-1 (GLP-1) secretion and short-chain fatty acid (SCFA) synthesis.	Improved insulin sensitivity and better metabolic regulation	Arshad <i>et al.</i> (2023)
<b>Whole-Food Plant-Based Diets</b>	Reduced systemic inflammatory activity and insulin-like growth factor-1 (IGF-1) signaling	Lower risk of cancer and metabolic disorders	Buffart <i>et al.</i> (2014)

Dietary fiber influences the composition of gut microbes by increasing production of short-chain fatty acids (SCFAs) that stabilize intestinal barrier integrity and reduce exposure to endotoxins causing inflammation. Moreover, moderate exercise increases insulin sensitivity and boosts mitochondrial activity concurrently. There is evidence from recommendations for cancer survivors that structured physical activity combined with dietary modifications can improve metabolic health and help to lower chronic inflammation. Moreover, there has been recent evidence to suggest that too much sugar can negatively affect brain plasticity, leading to neuroinflammation and reducing the levels of brain-derived neurotrophic factor (BDNF). Although such mechanisms can be influenced by dietary interventions, which can induce synaptic plasticity and brain resilience (Murphy *et al.*, 2025).

### Personalized medicine and precision nutrition

The interindividual variation in the metabolic results to dietary sugar underlines the importance of precision nutrition. Genetic variations that impact insulin signaling, lipid metabolism in the liver, and the release of inflammatory cytokines influence susceptibility to sugar-related diseases. Additionally, inflammation and glycemic swings are influenced by the gut microbiome. Metabolomics, microbiota profiling, and genetics are used in precision nutrition to personalize nutritional strategies (Arshad *et al.*, 2026). Through the assessment of glycemic responses, continuous glucose monitoring systems can facilitate.

**Table 11. Multi-Omics Approaches and Their Clinical Applications in Metabolic Health Assessment and Precision Nutrition**

<b>Biological Layer</b>	<b>Analytical Tool</b>	<b>Clinical Application</b>	<b>Reference</b>
<b>Genomics</b>	SNP Profiling	Finding the genetic predisposition to insulin resistance and more general metabolic problems	Arshad <i>et al.</i> (2023)
<b>Epigenomics</b>	DNA Methylation Clocks	Assessment of biological aging and epigenetic alterations associated with metabolic health	Park <i>et al.</i> (2017)
<b>Microbiomics</b>	Gut Microbiome Sequencing	Personalized probiotic and microbiota-targeted therapeutic interventions	Li <i>et al.</i> (2022)
<b>Metabolomics</b>	Postprandial Metabolic Analysis	Optimization of carbohydrate intake timing and metabolic response management	Arshad <i>et al.</i> (2023)

These strategies together study the personal metabolic issues of each individual. This could make nutritional therapies more effective. This could also prevent premature aging from happening too soon, or at least delay it.

### **Knowledge Gaps and Future Research Directions**

A diet high in sugar has been linked to a number of problems, including metabolic diseases and, in some cases, an acceleration of the aging process. I believe the evidence strongly suggests this, although there is still a lot that we don't know for sure at this point. The 2023 BMJ study, which is an umbrella review, addresses the substantial correlations between sugar intake and conditions like obesity, type 2 diabetes, and cardiac issues. However, in order to better determine whether sugar consumption is the direct cause of all these problems, and how much sugar is too much, we need these long-term randomized controlled trials. Otherwise, it seems like we're just guessing at the specifics here and there. The great bulk of the information that we do have access to at this point have come from observational studies and meta-analyses. Shortening—is yet not thoroughly studied. While metabolic factors are strongly emphasized in the guidelines for cancer survivors, it is not clear how sugar-induced insulin and inflammation factor into the promotion of tumor growth. Finally, there is a need for multi-omics clinical trials to validate personalized dietary advice, although the concept of precision nutrition models that combine genomes and metabolomics appears promising (Arshad *et al.*, 2026).

### **Conclusion**

Insulin resistance, oxidative stress, AGE accumulation, mitochondrial damage, and chronic inflammation are some of the mechanisms by which the overconsumption of sugar leads to metabolic diseases and premature aging. Vascular integrity, cognitive performance, immunity, and potentially the pathogenesis of cancer is all affected by the systemic consequences. Therefore, reducing sugar intake through personalized nutrition, targeted pharmaceutical approaches, and dietary strategies is an important molecular approach to facilitate healthy aging and enhance metabolic robustness.

### **Author Statement**

We, the authors, declare that the manuscript titled “**Molecular Mechanisms Linking Dietary Sugar to Metabolic Disorders, Inflammation, and Premature Aging**” is an original work and has not been submitted elsewhere for publication. The authors have all contributed significantly to the preparation of this review.

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### **Conflict of Interest Statement**

The authors declare no conflicts of interest related to this work.

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Al Mamun, A., Wu, Y., Monalisa, I., Jia, C., Zhou, K., Munir, F., Xiao, J., and Jia, J. (2021). Role of pyroptosis in spinal cord injury and its therapeutic implications. *Cells*, 10(2), Article 314. <https://doi.org/10.3390/cells10020314>

- Alam, Y. H., Kim, R., and Jang, C. (2022). Metabolism and health impacts of dietary sugars. *Journal of Lipid and Atherosclerosis*, 11(1), 20–38. <https://doi.org/10.12997/jla.2022.11.1.20>
- Aragno, M., and Mastrocola, R. (2017). Dietary sugars and endogenous formation of advanced glycation end products: Emerging mechanisms and implications. *Nutrients*, 9(4), Article 385. <https://doi.org/10.3390/nu9040385>
- Arshad, M. T., Ali, M. K. M., Awlqadr, F. H., Maqsood, S., Ikram, A., Hossain, M. S., Abdullahi, M. A., and Rashed, M. M. (2023). Mitigation of metabolic diseases through personalized nutrition: A critical in-depth review. *Journal of Food Science and Technology*, 60(11), 2733–2747. <https://doi.org/10.1007/s13197-023-05786-w> [Verify full details before submission]
- Augustin, L. S. A., Kendall, C. W. C., Jenkins, D. J. A., Willett, W. C., Astrup, A., Barclay, A. W., Björck, I., Brand-Miller, J. C., Brighenti, F., Buyken, A. E., Ceriello, A., La Vecchia, C., Livesey, G., Liu, S., Riccardi, G., Rizkalla, S. W., Sievenpiper, J. L., Trichopoulou, A., Wolever, T. M. S., . . . Poli, A. (2015). Glycemic index, glycemic load and glycemic response: An International Scientific Consensus Summit from the International Carbohydrate Quality Consortium (ICQC). *Nutrition, Metabolism and Cardiovascular Diseases*, 25(9), 795–815. <https://doi.org/10.1016/j.numecd.2015.05.005>
- Baharuddin, B. (2025). The metabolic and molecular mechanisms linking fructose consumption to lipogenesis and metabolic disorders. *Clinical Nutrition ESPEN*, 68, 1–11. <https://doi.org/10.1016/j.clnesp.2025.06.010>
- Baker, D. J., Wijshake, T., Tchkonja, T., LeBrasseur, N. K., Childs, B. G., van de Sluis, B., Kirkland, J. L., and van Deursen, J. M. (2011). Clearance of p16<sup>Ink4a</sup>-positive senescent cells delays ageing-associated disorders. *Nature*, 479(7372), 232–236. <https://doi.org/10.1038/nature10600>
- Bannister, A. J., and Kouzarides, T. (2011). Regulation of chromatin by histone modifications. *Cell Research*, 21(3), 381–395. <https://doi.org/10.1038/cr.2011.22>
- Basciano, H., Federico, L., and Adeli, K. (2005). Fructose, insulin resistance, and metabolic dyslipidemia. *Nutrition and Metabolism*, 2, Article 5. <https://doi.org/10.1186/1743-7075-2-5>
- Bhatti, J. S., Bhatti, G. K., and Reddy, P. H. (2017). Mitochondrial dysfunction and oxidative stress in metabolic disorders: A step towards mitochondria based therapeutic strategies. *Biochimica et Biophysica Acta (BBA) – Molecular Basis of Disease*, 1863(5), 1066–1077. <https://doi.org/10.1016/j.bbadis.2016.11.010>
- Blackburn, E. H., Epel, E. S., and Lin, J. (2015). Human telomere biology: A contributory and interactive factor in aging, disease risks, and protection. *Science*, 350(6265), 1193–1198. <https://doi.org/10.1126/science.aab3389>
- Bonomini, F., Rodella, L. F., and Rezzani, R. (2010). Metabolic syndrome, aging and involvement of oxidative stress. *Aging and Disease*, 1(2), 76–88.
- Brownlee, M. (2001). Biochemistry and molecular cell biology of diabetic complications. *Nature*, 414(6865), 813–820. <https://doi.org/10.1038/414813a>
- Buffart, L. M., Kalter, J., Sweegers, M. G., Courneya, K. S., Newton, R. U., Aaronson, N. K., Jacobsen, P. B., May, A. M., Galvão, D. A., Chinapaw, M. J. M., Steindorf, K., Spiegel, D., Irwin, M. L., Stuiver, M. M., Hayes, S., Griffith, K. A., Lucia, A., Mesters, I., van Weert, E., . . . Brug, J. (2017). Effects and moderators of exercise on quality of life and physical function in patients with cancer: An individual patient data meta-analysis of 34 RCTs. *Cancer Treatment Reviews*, 52, 91–104. <https://doi.org/10.1016/j.ctrv.2016.11.010> [Verify: 'Buffart and May' in text — confirm this is the intended source]
- Bzdok, D., Altman, N., and Krzywinski, M. (2018). Statistics versus machine learning. *Nature Methods*, 15(4), 233–234. <https://doi.org/10.1038/nmeth.4642>

- Cantó, C., and Auwerx, J. (2009). PGC-1 $\alpha$ , SIRT1 and AMPK, an energy sensing network that controls energy expenditure. *Current Opinion in Lipidology*, 20(2), 98–105. <https://doi.org/10.1097/MOL.0b013e328328d0a4>
- Capurso, C., and Capurso, A. (2012). From excess adiposity to insulin resistance: The role of free fatty acids. *Vascular Pharmacology*, 57(2–4), 91–97. <https://doi.org/10.1016/j.vph.2012.05.003>
- Chen, L., Chen, R., Wang, H., and Liang, F. (2015). Mechanisms linking inflammation to insulin resistance. *International Journal of Endocrinology*, 2015, Article 508409. <https://doi.org/10.1155/2015/508409>
- Chen, X., Li, C., and He, C. (2025). Oxidative stress in diabetes mellitus and its complications. *Chinese Medical Journal*, 138(1), 14–25. <https://doi.org/10.1097/CM9.0000000000003307> [Verify before submission]
- Clish, C. B. (2015). Metabolomics: An emerging but powerful tool for precision medicine. *Cold Spring Harbor Molecular Case Studies*, 1(1), Article a000588. <https://doi.org/10.1101/mcs.a000588>
- Coppé, J.-P., Desprez, P.-Y., Krtolica, A., and Campisi, J. (2010). The senescence-associated secretory phenotype: The dark side of tumor suppression. *Annual Review of Pathology: Mechanisms of Disease*, 5, 99–118. <https://doi.org/10.1146/annurev-pathol-121808-102144>
- Davey Smith, G., and Hemani, G. (2014). Mendelian randomization: Genetic anchors for causal inference in epidemiological studies. *Human Molecular Genetics*, 23(R1), R89–R98. <https://doi.org/10.1093/hmg/ddu328>
- Elliott, S. S., Keim, N. L., Stern, J. S., Teff, K., and Havel, P. J. (2002). Fructose, weight gain, and the insulin resistance syndrome. *The American Journal of Clinical Nutrition*, 76(5), 911–922. <https://doi.org/10.1093/ajcn/76.5.911>
- Epel, E. S., Blackburn, E. H., Lin, J., Dhabhar, F. S., Adler, N. E., Morrow, J. D., and Cawthon, R. M. (2004). Accelerated telomere shortening in response to life stress. *Proceedings of the National Academy of Sciences*, 101(49), 17312–17315. <https://doi.org/10.1073/pnas.0407162101>
- Fajstova, A., Galanova, N., Coufal, S., Malkova, J., Kostovcik, M., Cermakova, M., Pelantova, H., Kuzma, M., Sediva, B., and Hudcovic, T. (2020). Diet rich in simple sugars promotes pro-inflammatory response via gut microbiota alteration and TLR4 signaling. *Cells*, 9(12), Article 2701. <https://doi.org/10.3390/cells9122701>
- Ferraris, R. P. (2001). Dietary and developmental regulation of intestinal sugar transport. *Biochemical Journal*, 360(2), 265–276. <https://doi.org/10.1042/bj3600265>
- Fontana, L., Partridge, L., and Longo, V. D. (2010). Extending healthy life span—From yeast to humans. *Science*, 328(5976), 321–326. <https://doi.org/10.1126/science.1172539>
- Franceschi, C., Garagnani, P., Parini, P., Giuliani, C., and Santoro, A. (2018). Inflammaging: A new immune–metabolic viewpoint for age-related diseases. *Nature Reviews Endocrinology*, 14(10), 576–590. <https://doi.org/10.1038/s41574-018-0059-4>
- Giacco, F., and Brownlee, M. (2010). Oxidative stress and diabetic complications. *Circulation Research*, 107(9), 1058–1070. <https://doi.org/10.1161/CIRCRESAHA.110.223545>
- González, P., Lozano, P., Ros, G., and Solano, F. (2023). Hyperglycemia and oxidative stress: An integral, updated and critical overview of their metabolic interconnections. *International Journal of Molecular Sciences*, 24(11), Article 9352. <https://doi.org/10.3390/ijms24119352>
- Goodpaster, B. H., and Sparks, L. M. (2017). Metabolic flexibility in health and disease. *Cell Metabolism*, 25(5), 1027–1036. <https://doi.org/10.1016/j.cmet.2017.04.015>

- Hannou, S. A., Haslam, D. E., McKeown, N. M., and Herman, M. A. (2018). Fructose metabolism and metabolic disease. *Journal of Clinical Investigation*, 128(2), 545–555. <https://doi.org/10.1172/JCI96702>
- Hardie, D. G., Ross, F. A., and Hawley, S. A. (2012). AMPK: A nutrient and energy sensor that maintains energy homeostasis. *Nature Reviews Molecular Cell Biology*, 13(4), 251–262. <https://doi.org/10.1038/nrm3311>
- Hasin, Y., Seldin, M., and Lusic, A. (2017). Multi-omics approaches to disease. *Genome Biology*, 18, Article 83. <https://doi.org/10.1186/s13059-017-1215-1>
- Herman, M. A., and Birnbaum, M. J. (2021). Molecular aspects of fructose metabolism and metabolic disease. *Cell Metabolism*, 33(12), 2329–2354. <https://doi.org/10.1016/j.cmet.2021.09.010>
- Hotamisligil, G. S. (2017). Inflammation, metaflammation and immunometabolic disorders. *Nature*, 542(7640), 177–185. <https://doi.org/10.1038/nature21363>
- Huang, Y., Chen, Z., Chen, B., Li, J., Yuan, X., Li, J., Wang, W., Dai, T., Chen, H., Wang, Y., Wang, R., Wang, P., Guo, J., Dong, Q., Liu, C., Wei, Q., Cao, D., and Liu, L. (2023). Dietary sugar consumption and health: Umbrella review. *BMJ*, 381, Article e071609. <https://doi.org/10.1136/bmj-2022-071609>
- Hussain, S. A., Al-Nimer, M. S. M., Al-Anbari, H. H., and Hameed, H. G. (2026). DNA methylation and personalized nutrition: Mechanisms and implications. *International Journal of Molecular Sciences*, 27(2), Article 566. [Advance online publication — verify DOI and final details before submission]
- Integrative HMP (iHMP) Research Network Consortium. (2019). The integrative human microbiome project. *Nature*, 569(7758), 641–648. <https://doi.org/10.1038/s41586-019-1238-8>
- Johnson, R. J., Nakagawa, T., Sánchez-Lozada, L. G., Shafiu, M., Sundaram, S., Le, M., Ishimoto, T., Sautin, Y. Y., and Lanaspá, M. A. (2013a). Sugar, uric acid, and the etiology of diabetes and obesity. *Diabetes*, 62(10), 3307–3315. <https://doi.org/10.2337/db12-1814>
- Johnson, R. J., Sánchez-Lozada, L. G., Andrews, P., and Lanaspá, M. A. (2017). Perspective: A historical and scientific perspective of sugar and its relation with obesity and diabetes. *Advances in Nutrition*, 8(3), 412–422. <https://doi.org/10.3945/an.116.014654>
- Kaszycki, J., and Kim, M. (2025). Epigenetic regulation of transcription factors involved in NLRP3 inflammasome and NF-κB signaling pathways. *Frontiers in Immunology*, 16, Article 1529756. <https://doi.org/10.3389/fimmu.2025.1529756>
- Koh, A., De Vadder, F., Kovatcheva-Datchary, P., and Bäckhed, F. (2016). From dietary fiber to host physiology: Short-chain fatty acids as key bacterial metabolites. *Cell*, 165(6), 1332–1345. <https://doi.org/10.1016/j.cell.2016.05.041>
- Lanaspá, M. A., Sánchez-Lozada, L. G., Cicerchi, C., Li, N., Roncal-Jimenez, C. A., Ishimoto, T., Le, M., Garcia, G. E., Thomas, J. B., Rivard, C. J., Andres-Hernando, A., Hunter, B., Schreiner, G., Rodriguez-Iturbe, B., Sautin, Y. Y., and Johnson, R. J. (2012). Uric acid stimulates fructokinase and accelerates fructose metabolism in the development of fatty liver. *PLOS ONE*, 7(10), Article e47948. <https://doi.org/10.1371/journal.pone.0047948>
- Laplante, M., and Sabatini, D. M. (2012). mTOR signaling in growth control and disease. *Cell*, 149(2), 274–293. <https://doi.org/10.1016/j.cell.2012.03.017>
- Li, D., Li, Y., Yang, S., Lu, J., Jin, X., and Wu, M. (2022). Diet–gut microbiota–epigenetics in metabolic diseases: From mechanisms to therapeutics. *Biomedicine and Pharmacotherapy*, 153, Article 113290. <https://doi.org/10.1016/j.biopha.2022.113290>
- Li, X., Zhang, Y., Wang, Q., Chen, H., and Liu, J. (2025). Integrative multi-omics analysis of sleep apnea: Gut microbiome, metabolomics, proteomics, and

- phenome-wide association study. *Frontiers in Genetics*, *16*, Article 1512847. [Verify full details before submission]
- López-Otín, C., Blasco, M. A., Partridge, L., Serrano, M., and Kroemer, G. (2013). The hallmarks of aging. *Cell*, *153*(6), 1194–1217. <https://doi.org/10.1016/j.cell.2013.05.039>
- Ludwig, D. S. (2002). The glycemic index: Physiological mechanisms relating to obesity, diabetes, and cardiovascular disease. *JAMA*, *287*(18), 2414–2423. <https://doi.org/10.1001/jama.287.18.2414>
- Lustig, R. H., Schmidt, L. A., and Brindis, C. D. (2012). The toxic truth about sugar. *Nature*, *482*(7383), 27–29. <https://doi.org/10.1038/482027a>
- Ma, X., Nan, F., Liang, H., Shu, P., Fan, X., Song, X., Hou, Y., and Zhang, D. (2022). Excessive intake of sugar: An accomplice of inflammation. *Frontiers in Immunology*, *13*, Article 988481. <https://doi.org/10.3389/fimmu.2022.988481>
- Macdonald, I. A. (2016). A review of recent evidence relating to sugars, insulin resistance and diabetes. *European Journal of Nutrition*, *55*(Suppl. 2), 17–23. <https://doi.org/10.1007/s00394-016-1340-8>
- Malik, V. S., Popkin, B. M., Bray, G. A., Després, J.-P., Willett, W. C., and Hu, F. B. (2010). Sugar-sweetened beverages and risk of metabolic syndrome and type 2 diabetes: A meta-analysis. *Diabetes Care*, *33*(11), 2477–2483. <https://doi.org/10.2337/dc10-1079>
- Manning, B. D., and Cantley, L. C. (2007). AKT/PKB signaling: Navigating downstream. *Cell*, *129*(7), 1261–1274. <https://doi.org/10.1016/j.cell.2007.06.009>
- Menni, C., Zhai, G., MacGregor, A., Prehn, C., Römisch-Margl, W., Suhre, K., Adamski, J., Cassidy, A., Illig, T., Spector, T. D., and Valdes, A. M. (2013). Targeted metabolomics profiles are strongly correlated with nutritional patterns in women. *Metabolomics*, *9*(2), 506–514. <https://doi.org/10.1007/s11306-012-0469-6>
- Mizushima, N., and Komatsu, M. (2011). Autophagy: Renovation of cells and tissues. *Cell*, *147*(4), 728–741. <https://doi.org/10.1016/j.cell.2011.10.026>
- Monnier, V. M., Sell, D. R., Genuth, S., and DCCT Research Group. (2005). Glycation products as markers and predictors of the progression of diabetic complications. *Annals of the New York Academy of Sciences*, *1043*(1), 567–581. <https://doi.org/10.1196/annals.1333.065>
- Moreno-Fernández, S., Garcés-Rimón, M., Vera, G., Astier, J., Landrier, J.-F., and Miguel, M. (2023). High fat and high glucose diet-induced metabolic disorders in C57BL/6J mice: Impact of obesity and oxidative stress. *Nutrients*, *15*(4), Article 1023. <https://doi.org/10.3390/nu15041023>
- Moreno-Fernández, S., Garcés-Rimón, M., Vera, G., Astier, J., Landrier, J.-F., and Miguel, M. (2018). High fat/high glucose diet induces metabolic syndrome in an experimental rat model. *Nutrients*, *10*(10), Article 1483. <https://doi.org/10.3390/nu10101483> [Verify: confirm this is the 2018 source cited in the Metabolomics section]
- Murphy, T., Dias, G. P., and Thuret, S. (2014). Effects of diet on brain plasticity in animal and human studies: Mind the gap. *Neural Plasticity*, *2014*, Article 563160. <https://doi.org/10.1155/2014/563160> [Verify: article cites Murphy *et al.* 2025 — confirm correct year and source]
- Nicoletti, C. F., Delfino, H. B. P., Ferreira, F. C., Pinhel, M. A. S., and Nonino, C. B. (2024). Non-coding RNAs and DNA methylation in metabolic diseases. *Frontiers in Genetics*, *15*, Article 1378327. [Verify full details before submission]
- Nishikawa, T., Edelstein, D., Du, X. L., Yamagishi, S., Matsumura, T., Kaneda, Y., Yorek, M. A., Beebe, D., Oates, P. J., Hammes, H.-P., Giardino, I., and Brownlee, M. (2000). Normalizing mitochondrial superoxide production

- blocks three pathways of hyperglycaemic damage. *Nature*, 404(6779), 787–790. <https://doi.org/10.1038/35008121>
- O'Connor, L., Imamura, F., Lentjes, M. A. H., Khaw, K.-T., Wareham, N. J., and Forouhi, N. G. (2018). Prospective associations and population impact of sweet beverage intake and type 2 diabetes, and effects of substitutions with alternative beverages. *Clinical Nutrition*, 37(1), 181–189. <https://doi.org/10.1016/j.clnu.2016.12.009>
- Park, J. H., Yoo, Y., and Park, Y. J. (2017). Epigenetics: Connecting environment and genotype to phenotype and disease. *Preventive Nutrition and Food Science*, 22(2), 81–89. <https://doi.org/10.3746/pnf.2017.22.2.81>
- Paul, R. G., and Bailey, A. J. (1996). Glycation of collagen: The basis of its central role in the late complications of ageing and diabetes. *International Journal of Biochemistry and Cell Biology*, 28(12), 1297–1310. [https://doi.org/10.1016/S1357-2725\(96\)00079-9](https://doi.org/10.1016/S1357-2725(96)00079-9)
- Petersen, M. C., and Shulman, G. I. (2018). Mechanisms of insulin action and insulin resistance. *Physiological Reviews*, 98(4), 2133–2223. <https://doi.org/10.1152/physrev.00063.2017>
- Popkin, B. M., and Hawkes, C. (2016). Sweetening of the global diet, particularly beverages: Patterns, trends, and policy responses. *The Lancet Diabetes and Endocrinology*, 4(2), 174–186. [https://doi.org/10.1016/S2213-8587\(15\)00419-2](https://doi.org/10.1016/S2213-8587(15)00419-2)
- Rehman, K., and Akash, M. S. H. (2016). Mechanisms of inflammatory responses and development of insulin resistance: How are they interlinked? *Journal of Biomedical Science*, 23, Article 87. <https://doi.org/10.1186/s12929-016-0303-y> [Verify: cited as Rehman and Akash (2020) in Table 2 — confirm correct year]
- Rippe, J. M., and Angelopoulos, T. J. (2016). Relationship between added sugars consumption and chronic disease risk factors: Current understanding. *Nutrients*, 8(11), Article 697. <https://doi.org/10.3390/nu8110697>
- Rorsman, P., and Ashcroft, F. M. (2018). Pancreatic  $\beta$ -cell electrical activity and insulin secretion: Of mice and men. *Physiological Reviews*, 98(1), 117–214. <https://doi.org/10.1152/physrev.00008.2017>
- Rutledge, A. C., and Adeli, K. (2007). Fructose and the metabolic syndrome: Pathophysiology and molecular mechanisms. *Nutrition Reviews*, 65(6, Suppl. 1), S13–S23. <https://doi.org/10.1111/j.1753-4887.2007.tb00332.x>
- Saltiel, A. R., and Kahn, C. R. (2001). Insulin signalling and the regulation of glucose and lipid metabolism. *Nature*, 414(6865), 799–806. <https://doi.org/10.1038/414799a>
- Samuel, V. T., and Shulman, G. I. (2012). Mechanisms for insulin resistance: Common threads and missing links. *Cell*, 148(5), 852–871. <https://doi.org/10.1016/j.cell.2012.02.017>
- Schmidt, A. M., Yan, S. D., Yan, S. F., and Stern, D. M. (1999). The multiligand receptor RAGE as a progression factor amplifying immune and inflammatory responses. *Journal of Clinical Investigation*, 108(7), 949–955. <https://doi.org/10.1172/JCI14002>
- Schmidt, A. M., Hori, O., and Stern, D. M. (2001). RAGE: A novel cellular receptor for advanced glycation end products. *Diabetes*, 45(Suppl. 3), S77–S80. <https://doi.org/10.2337/diab.45.3.S77>
- Semchyshyn, H. M., Miedzobrodzki, J., and Bayliak, M. M. (2024). AGE–RAGE axis in metabolic disorders and aging. *Frontiers in Nutrition*, 11, Article 1289452. <https://doi.org/10.3389/fnut.2024.1289452>
- Shi, Y. (2021). Fructose and metabolic diseases: New findings, new questions. *Nutrition*, 89, Article 111232. <https://doi.org/10.1016/j.nut.2021.111232>

- Shi, Y.-N., Li, Y., Wang, H., and Zhang, J. (2021). Fructose consumption and metabolic diseases: Too much to be good. *Chinese Medical Journal*, *134*(11), 1276–1285. <https://doi.org/10.1097/CM9.0000000000001546>
- Singh, R., Barden, A., Mori, T., and Beilin, L. (2001). Advanced glycation end-products: A review. *Diabetologia*, *44*(2), 129–146. <https://doi.org/10.1007/s001250051591>
- Softic, S., Cohen, D. E., and Kahn, C. R. (2016). Role of dietary fructose and hepatic de novo lipogenesis in fatty liver disease. *Digestive Diseases and Sciences*, *61*(5), 1282–1293. <https://doi.org/10.1007/s10620-016-4054-0> [Verify: article cites Softic *et al.* 2020 — confirm correct year]
- Sonnenburg, J. L., and Bäckhed, F. (2016). Diet–microbiota interactions as moderators of human metabolism. *Nature*, *535*(7610), 56–64. <https://doi.org/10.1038/nature18846>
- Sousa, E. S., Oliveira, A. G., Silva, M. C., and Monteiro, M. P. (2023). High glucose modulates macrophage function and inflammatory responses. *Frontiers in Immunology*, *14*, Article 1130662. <https://doi.org/10.3389/fimmu.2023.1130662>
- Srour, B., Fezeu, L. K., Kesse-Guyot, E., Allès, B., Méjean, C., Andrianasolo, R. M., Chazelas, E., Deschasaux, M., Hercberg, S., Galan, P., Monteiro, C. A., Julia, C., and Touvier, M. (2019). Ultra-processed food intake and risk of cardiovascular disease: Prospective cohort study. *BMJ*, *365*, Article 11451. <https://doi.org/10.1136/bmj.11451>
- Stanhope, K. L. (2016). Sugar consumption, metabolic disease and obesity: The state of the controversy. *Critical Reviews in Clinical Laboratory Sciences*, *53*(1), 52–67. <https://doi.org/10.3109/10408363.2015.1084990>
- Suarez, G., Rajaram, R., Oronsky, A. L., and Gawinowicz, M. A. (1989). Nonenzymatic glycation of bovine serum albumin by fructose (fructation): Comparison with the Maillard reaction initiated by glucose. *Journal of Biological Chemistry*, *264*(7), 3674–3679. [https://doi.org/10.1016/S0021-9258\(18\)83697-4](https://doi.org/10.1016/S0021-9258(18)83697-4)
- Taskinen, M.-R., Packard, C. J., and Borén, J. (2019). Dietary fructose and the metabolic syndrome. *Nutrients*, *11*(9), Article 1987. <https://doi.org/10.3390/nu11091987>
- Tappy, L., and Lê, K.-A. (2010). Metabolic effects of fructose and the worldwide increase in obesity. *Physiological Reviews*, *90*(1), 23–46. <https://doi.org/10.1152/physrev.00019.2009>
- Tchkonina, T., Zhu, Y., van Deursen, J., Campisi, J., and Kirkland, J. L. (2013). Cellular senescence and the senescent secretory phenotype: Therapeutic opportunities. *Journal of Clinical Investigation*, *123*(3), 966–972. <https://doi.org/10.1172/JCI64098>
- Te Morenga, L., Mallard, S., and Mann, J. (2013). Dietary sugars and body weight: Systematic review and meta-analyses of randomised controlled trials and cohort studies. *BMJ*, *346*, Article e7492. <https://doi.org/10.1136/bmj.e7492>
- Tratnjek, L., Kreft, M. E., and Živin, M. (2026). Sugar, inflammation, and cancer: Emerging molecular mechanisms. *Frontiers in Immunology*, *17*, Article 1731784. [Advance online publication — verify DOI and final details before submission]
- Turnbaugh, P. J., Ley, R. E., Hamady, M., Fraser-Liggett, C. M., Knight, R., and Gordon, J. I. (2007). The human microbiome project. *Nature*, *449*(7164), 804–810. <https://doi.org/10.1038/nature06244>
- Uribarri, J., Woodruff, S., Goodman, S., Cai, W., Chen, X., Pyzik, R., and Vlassara, H. (2010). Advanced glycation end products in foods and a practical guide to their reduction in the diet. *Journal of the American Dietetic Association*, *110*(6), 911–916. <https://doi.org/10.1016/j.jada.2010.03.018>

- Vasudevan, A., Yu, Y., Bhatt, D. L., and Bhatt, A. B. (2024). Fructose-mediated remodeling of liver-resident Kupffer cells in metabolic disease. *Hepatology Communications*, 8(4), Article e0393. <https://doi.org/10.1097/HC9.0000000000000393> [Verify: confirm this matches the Vasudevan *et al.* 2024 cited in the text]
- Verzija, N., DeGroot, J., Thorpe, S. R., Bank, R. A., Shaw, J. N., Lyons, T. J., Bijlsma, J. W. J., Lafeber, F. P. J. G., Baynes, J. W., and TeKoppele, J. M. (2000). Effect of collagen turnover on the accumulation of advanced glycation end products. *Journal of Biological Chemistry*, 275(50), 39027–39031. <https://doi.org/10.1074/jbc.M006700200>
- Vlassara, H., and Uribarri, J. (2014). Advanced glycation end products (AGE) and diabetes: Cause, effect, or both? *Current Diabetes Reports*, 14, Article 453. <https://doi.org/10.1007/s11892-013-0453-1>
- von Zglinicki, T. (2002). Oxidative stress shortens telomeres. *Trends in Biochemical Sciences*, 27(7), 339–344. [https://doi.org/10.1016/S0968-0004\(02\)02110-2](https://doi.org/10.1016/S0968-0004(02)02110-2)
- Vousden, K. H., and Prives, C. (2009). Blinded by the light: The growing complexity of p53. *Cell*, 137(3), 413–431. <https://doi.org/10.1016/j.cell.2009.04.037>
- Wallace, D. C. (2005). A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: A dawn for evolutionary medicine. *Annual Review of Genetics*, 39, 359–407. <https://doi.org/10.1146/annurev.genet.39.110304.095751>
- Wang, Q., and Wu, H. (2018). T cells in adipose tissue: Critical players in immunometabolism. *Frontiers in Immunology*, 9, Article 2509. <https://doi.org/10.3389/fimmu.2018.02509>
- Wang, Z., Li, Q., Li, H., Yang, C., Zhang, Y., and Zhao, Y. (2025). Multi-omics integration reveals the impact of Mediterranean diet on hepatic metabolism and gut microbiota in mice with metabolic dysfunction-associated steatotic liver disease. *Frontiers in Nutrition*, 12, Article 1644014. <https://doi.org/10.3389/fnut.2025.1644014>
- Wellen, K. E., Hatzivassiliou, G., Sachdeva, U. M., Bui, T. V., Cross, J. R., and Thompson, C. B. (2009). ATP-citrate lyase links cellular metabolism to histone acetylation. *Science*, 324(5930), 1076–1080. <https://doi.org/10.1126/science.1164097>
- Wishart, D. S. (2016). Emerging applications of metabolomics in drug discovery and precision medicine. *Nature Reviews Drug Discovery*, 15(7), 473–484. <https://doi.org/10.1038/nrd.2016.32>
- Younossi, Z. M., Koenig, A. B., Abdelatif, D., Fazel, Y., Henry, L., and Wymer, M. (2016). Global epidemiology of nonalcoholic fatty liver disease—Meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology*, 64(1), 73–84. <https://doi.org/10.1002/hep.28431>
- Zhang, L., Li, X., Wang, Y., Chen, Z., and Liu, J. (2024). Microbiome remodeling during aging and metabolic dysfunction. *Frontiers in Genetics*, 15, Article 1450064. <https://doi.org/10.3389/fgene.2024.1450064>
- Zhao, J., Miao, K., Wang, H., Ding, H., and Wang, D. W. (2013). Association between telomere length and type 2 diabetes mellitus: A meta-analysis. *Diabetes*, 63(1), 354–362. <https://doi.org/10.2337/db13-0246>
- Zhao, S., Xu, W., Jiang, W., Yu, W., Lin, Y., Zhang, T., Yao, J., Zhou, L., Zeng, Y., Li, H., Li, Y., Shi, J., An, W., Hancock, S. M., He, F., Qin, L., Chin, J., Yang, P., Chen, X., . . . Zhao, Y. (2010). Regulation of cellular metabolism by protein lysine acetylation. *Science*, 327(5968), 1000–1004. <https://doi.org/10.1126/science.1179689>